

Dopamine and Oxytocin Interactions Underlying Behaviors: Potential Contributions to Behavioral Disorders

Tracey A. Baskerville¹ & Alison J. Douglas²

¹ Division of Clinical Neurosciences, Gartscube Estate, University of Glasgow, UK

² Centre for Integrative Physiology, University of Edinburgh, UK

Keywords

Autism; Bonding; Depression; Parental brain; Sexual dysfunction; Social disorders.

Correspondence

Dr. Tracey Baskerville, Division of Clinical Neurosciences, University of Glasgow, Gartscube Estate, Bearsden Road, Glasgow G12 8QQ
Tel.: (141) 330-7103;
Fax: (141) 943-0215;
E-mail: tab3b@clinmed.gla.ac.uk

doi: 10.1111/j.1755-5949.2010.00154.x

Dopamine is an important neuromodulator that exerts widespread effects on the central nervous system (CNS) function. Disruption in dopaminergic neurotransmission can have profound effects on mood and behavior and as such is known to be implicated in various neuropsychiatric behavioral disorders including autism and depression. The subsequent effects on other neurocircuits due to dysregulated dopamine function have yet to be fully explored. Due to the marked social deficits observed in psychiatric patients, the neuropeptide, oxytocin is emerging as one particular neural substrate that may be influenced by the altered dopamine levels subserving neuropathologic-related behavioral diseases. Oxytocin has a substantial role in social attachment, affiliation and sexual behavior. More recently, it has emerged that disturbances in peripheral and central oxytocin levels have been detected in some patients with dopamine-dependent disorders. Thus, oxytocin is proposed to be a key neural substrate that interacts with central dopamine systems. In addition to psychosocial improvement, oxytocin has recently been implicated in mediating mesolimbic dopamine pathways during drug addiction and withdrawal. This bi-directional role of dopamine has also been implicated during some components of sexual behavior. This review will discuss evidence for the existence dopamine/oxytocin positive interaction in social behavioral paradigms and associated disorders such as sexual dysfunction, autism, addiction, anorexia/bulimia, and depression. Preliminary findings suggest that whilst further rigorous testing has to be conducted to establish a dopamine/oxytocin link in human disorders, animal models seem to indicate the existence of broad and integrated brain circuits where dopamine and oxytocin interactions at least in part mediate socio-affiliative behaviors. A profound disruption to these pathways is likely to underpin associated behavioral disorders. Central oxytocin pathways may serve as a potential therapeutic target to improve mood and socio-affiliative behaviors in patients with profound social deficits and/or drug addiction.

Introduction

The neurobiological and neurochemical mechanisms underlying the cause of prevalent psychiatric behavioral disorders such as autism in humans are not yet fully elucidated. In addition to contributing factors, including genetic predisposition and psychosocial environment, disruption to major central nervous system (CNS)

neurotransmitter pathways largely influences the onset of psychiatric disorders in patients. Current therapeutic interventions are pharmacological agents, which alleviate symptoms or redress brain neurotransmitter imbalance (often coupled with psychotherapeutic approaches such as cognitive behavioral therapy). Of the many central neurotransmitters believed to be implicated in CNS behavioral disorders, the monoamine, dopamine has

received much attention due to its extensive innervation of the brain, widespread receptor distribution and subsequent role across a broad spectrum of central functions and behaviors such as cognition, emotion, perception, motivation, reward, and sleep, in addition to peripheral actions on the cardiovascular and renal systems. Disturbances in central dopaminergic pathways are known pathologic mechanisms contributing to major psychiatric illnesses such as Parkinson's disease and schizophrenia. However, such dopaminergic disruptions are also believed to underpin several behavioral disorders including social anxiety, major depressive disorders and compulsive behaviors [1–3]. Here we intend to give an overview of the role of dopamine in selected behaviors and associated disorders. Although a role for dopamine in some behaviors such as sexual dysfunction is highly likely, for others its effects and the neurocircuitries it employs remain to be fully elucidated. However, the neuropeptide oxytocin is one central mediator in particular that is garnering much research interest due to its widespread effects on CNS function.

Oxytocin has a classical role in endocrine regulation where it acts as an important mediator in parturition and the milk ejection reflex during lactation [4]. Beyond its involvement in endocrine function, oxytocin acts in the brain as a key substrate for a range of social behaviors (including social bonding, parental behavior and sexual behavior and nonsocial behaviors such as stress, anxiety and aggression) [5]. The influential role of oxytocin in mediating social behavior is due to its widespread projections and receptor distribution the patterns of which determine behavior quality. Neurologic behavioral disorders caused by profound disruptions to key dopaminergic pathways in the brain are known to adversely affect prosocial behavior in mammals [6–8]. Thus, it is not surprising that oxytocin has been potentially implicated in several dopamine-dependent behavioral disorders including anxiety and autism and as such is emerging as a potential therapeutic target in the treatment of these diseases.

While the relationship between central dopamine and oxytocin is evident in some preclinical studies investigating sexual and social behavior [9–11], evidence of neural crosstalk between these two systems under pathophysiological conditions is underresearched. This review will focus on findings from preclinical and clinical (where possible) studies which have attempted to delineate a dopaminergic–oxytocinergic link and highlight potential treatment options in the following behavioral disorders: sexual dysfunction, autism, addiction, depression, and anorexia/bulimia. First, we will outline dopamine and oxytocin sources and targets and their roles in selected behaviors. This will clarify potential brain regions

involved and relevant modes of interaction. We will then explain the dopamine–oxytocin interaction in sexual function and dysfunction before analysing their interaction and role in potential dysfunction in other selected social and nonsocial contexts.

Dopamine

Dopamine Synthesis and Distribution

Dopamine is an immensely important central neurotransmitter that has widespread projections and functions throughout the CNS. Dopamine synthesis is a two-step reaction and involves the creation of L-dihydroxyphenylalanine (L-DOPA) from L-tyrosine via tyrosine hydroxylase. L-DOPA is then converted to dopamine by DOPA decarboxylase. Dopamine is then enzymatically converted to 3,4-dihydroxyphenyl acetic acid (DOPAC) and 3-methoxytyramine (3-MT) via the enzymes monoamine oxidase (MAO) and catechol-O-methyl transferase (COMT), respectively. Finally, DOPA and 3-MT are further degraded by COMT and MAO, respectively, to yield the inactive homovanillic acid (HVA)

Dopamine Release

Dopamine has a key role in a range of neurochemical and neurohormonal functions including cognition, sexual behavior, milk production, arousal, reward, coordination and motricity. Dopaminergic neuronal cell bodies originating in the substantia nigra (SN), hypothalamus, ventral tegmental area (VTA), arcuate nucleus and the zona incerta project to various brain structures and comprise six main pathways summarized in Figure 1 and their functions in Table 2. The nigrostriatal pathway originates in the SN and projects to the striatum where it controls the initiation and movement of muscle via the prefrontal cortex. Mesolimbic pathway cell bodies are found in the VTA and terminate in various limbic regions such as the nucleus accumbens (NA) and amygdala, where they are involved in reward, desire and reinforcement behaviors. Mesocortical dopamine fibers originate in the same region but project to the cortex where they mediate emotional and motivational responses. The tuberoinfundibular dopamine system has cell bodies in the arcuate nucleus and periventricular region of the hypothalamus where they project to the median eminence to regulate anterior pituitary prolactin secretion. The hypothalamic-derived incerto-hypothalamic dopamine pathway innervates the dorsal anterior hypothalamus, including the supraoptic nucleus (SON) and paraventricular nucleus (PVN) and the lateral septal nuclei where it is believed to have a role in endocrine regulation and sexual behavior [12,13].

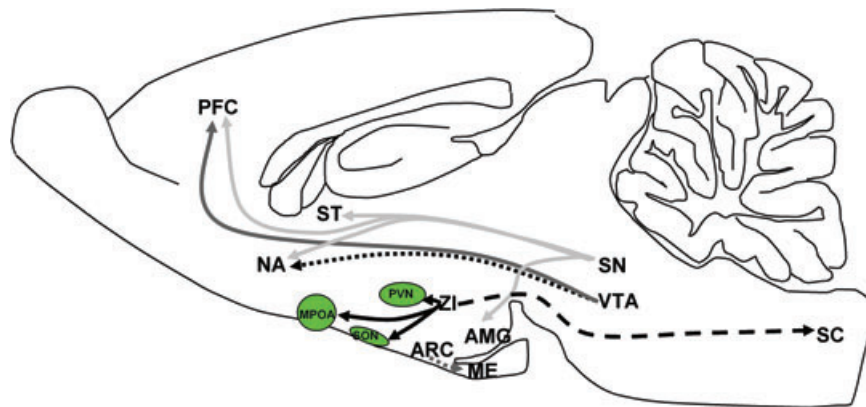


Figure 1 Major dopamine pathways in the rat brain. The nigrostriatal pathways are comprised of dopamine cell bodies in the SN, from here dopamine fibers innervate several brain regions including the ST, PFC, NA, and AMG (light gray line). Mesocortical and mesolimbic dopamine pathways originate in the VTA and project to the PFC (dark gray line), and NA (black-dotted line), respectively. The tuberoinfundibular dopamine system is comprised of dopamine fibers originating in the ARC and terminating in the ME (dark gray dotted line). Dopamine projections from the ZI to the

MPOA, SON, and PVN of the hypothalamus comprise the incerto-hypothalamic dopamine pathway (black line). The diencephalospinal dopamine system originates in the hypothalamus and projects to the thoracolumbar spinal cord (black-dashed line). PFC, prefrontal cortex; NA, nucleus accumbens; ZI, zona incerta; MPOA, medial preoptic nucleus; PVN, paraventricular nucleus; SON, supraoptic nucleus; AMG, amygdala; ARC, arcuate nucleus; VTA, ventral tegmental area; ME, median eminence; ST, striatum; SC, spinal cord.

Finally, the diencephalospinal dopamine system originates in the hypothalamus and projects to the thoracic and lumbar spinal cord where it has a role in spinal reflex functions such as the stretch reflex [14,15] and may also contribute to spinal control of penile erection [16].

Dopamine Receptors

Five dopamine receptors exist in the CNS and comprise D1-like (D1 and D5) and D2-like (D2, D3, and D4) receptor subgroups. The receptors can be divided into two separate subgroups depending on the transduction system to which they are coupled to (1) D1-like receptors (D1 and D5) which positively activate adenylate cyclase and (2) D2-like receptors (D2, D3, and D4) which are negatively or not coupled to the enzyme. There is generally widespread expression of all dopamine receptors in the brain with abundant levels of D1 and D2 receptors and moderate expression of D3, D4, and D5 receptors [17–21]. D1 and D2 receptors are found in the striatum, cortex, hypothalamus, olfactory bulbs, and SN [19,22]. D3 receptor expression is more restricted, with the NA, olfactory tubercles and the Islands of Calleja possessing moderate to high levels of the D3 receptor [19]. In comparison to D2 receptors, D4 receptor levels appear to be less abundant in subcortical structures. The cortex, hippocampus, and striatum have all been shown to possess D4 receptors [18,19]. Finally, D5 receptor expression in the rat brain is comparatively scarce, however, D5 recep-

tors have been shown to exist in the striatum, cortex, substantia nigra pars compacta, and NA [20].

Oxytocin

Oxytocin is a classical neuroendocrine neurohypophysial hormone, but over the last 20 years it has emerged as an influential hormone released in the brain which can initiate a wide spectrum of central effects in both males and females (see Table 1).

In the brain, the actions of oxytocin have been shown to be important in coordinating well-defined activities related to socio-sexual behaviors. Oxytocin pathways subserving maternal and social behavior are believed to be important in governing familial and nonfamilial bonds [23–26]. Another key, sometimes overlooked, role is in appetite-related behaviors where oxytocin both centrally and peripherally restrains food intake and decreases blood osmolality [27,28]. Furthermore, central oxytocin has also been shown to have anxiolytic and antistress properties whereby oxytocin-treated mice engage in more risk-taking, explorative, and investigative behaviors [29–31].

Oxytocin Synthesis and Distribution

Oxytocin is primarily synthesized in hypothalamic magnocellular neurosecretory cells in the SON and PVN, where it is transported to the neurohypophysis and released into the blood. From here oxytocin has a

Table 1 Oxytocin systems

Oxytocin systems	Origin	Projections	Function
Magnocellular (axonal)	SON/PVN	Posterior pituitary	Parturition, uterine contractions, milk ejection reflex
Magnocellular (dendritic) Parvocellular	SON/PVN PVN	SON/PVN, extrahypothalamic regions Ventral tegmental area, hippocampus, brainstem, spinal cord	Autoregulation, endocannabinoid stimulation Penile erection, ejaculation, gastric reflexes, respiration

Central oxytocin pathways. Within the CNS, two major oxytocin pathways exist; (1) the magnocellular oxytocin system originating in the SON and PVN can be further subdivided by its release characteristics in to axonal release (into the posterior pituitary) which regulates reproductive behavior and dendritic release (within the SON and PVN and may diffuse to other distant sites) to mediate oxytocin autoregulation (2) the parvocellular oxytocin system originates in the parvocellular PVN and projects to numerous CNS sites to regulate autonomic functions such as respiration and gastric reflexes.

vital role in reproduction, mediating smooth muscle contraction at precisely defined times to facilitate delivery of uterine contents at birth and milk (nutrition) to offspring during suckling [27,32–34]. Similarly, oxytocin is also generated in parvocellular neurons in the PVN, which project to extrahypothalamic regions within the CNS where they have a role in mediating various autonomic functions [35–38]. Oxytocinergic fibers are not restricted to the hypothalamus but also lie in various other brain regions including the hippocampus, cortex, SN, brain stem, and the spinal cord [4,39,40]. With its diffuse potential targets, oxytocin is able to influence a range of neuroendocrine-mediated functions governing social and affiliative behaviors such as maternal and socio-sexual behavior [9,26,32,41–44].

Oxytocin Release

As previously mentioned, magnocellular and parvocellular oxytocin release into systemic circulation and the CNS occurs via projections to the posterior pituitary and extrahypothalamic brain regions, respectively (see Table 2). Oxytocin release from axon terminals occurs in the classical manner where axonal terminal release is preceded by

an influx of calcium into axonal terminals in response to an invading action potential. However, as first demonstrated by Moos et al. [45], oxytocin can also be released somatodendritically from magnocellular oxytocin neurons in the PVN and SON to regulate its own release. This finding was further substantiated in numerous *in vivo* studies using microdialysis to quantitatively measure oxytocin release in the plasma and the brain of parturient and lactating rats [46–48]. Unlike axonal release of oxytocin, dendritic release of oxytocin is triggered by release of calcium from intracellular stores and is generally electrically independent [49,50]. Central (axon terminal) and peripheral (via hypophyseal secretion into circulation) oxytocin release from magnocellular cells can act synergistically to influence behavioral consequences. During various paradigms like suckling, there is a concomitant release of oxytocin into the bloodstream, SON, and PVN [46,51]. Such synergy between the central and peripheral oxytocin systems does not always exist and there can be an apparent disassociation between the two as seen during a psychosocial stressor such as social defeat [52,53]. Engelmann et al. [54] demonstrated that whilst intra-SON oxytocin release increased in response to social defeat, peripheral oxytocin release remained unaffected.

Table 2 Dopamine systems

Dopamine systems	Origin	Projections	Function
Nigrostriatal	SN (A9)	Striatum	Motricity
Mesocortical	VTA(A10)	Cortex	Emotionality
Mesolimbic	VTA(A10)	NAC	Reward and desire
Tuberoinfundibular	Arcuate Nucleus (A12)	Median Eminence	Regulation of prolactin release
Incertohypothalamic	Zona Incerta (A13) Periventricular region (A14)	Various hypothalamic nuclei, thalamus	Sexual arousal and copulation
Diencephalospinal	Hypothalamus (A11)	Spinal cord	Afferent stretch reflex, contraction of penile striated muscles

Central dopamine pathways. Within the CNS, three major dopamine pathways exist; the nigrostriatal, mesocortical/mesolimbic and tuberoinfundibular systems which influence motor function, mood, reward and neuropeptide release. There are two additional minor dopamine pathways; the incertohypothalamic and diencephalospinal systems which are believed to modulate elements of sexual behavior. SN = substantia nigra; VTA = ventral tegmental area; Nac = nucleus accumbens).

Thus, it can be seen that during certain neuroendocrine-mediated behaviors, centrally acting and peripherally acting oxytocin may act in unison or independently to exert their behaviorally specific effects.

Oxytocin Receptors

The encoded oxytocin receptor is a 389-amino acid polypeptide with seven transmembrane domains and is thus part of the G protein-coupled receptor family. When oxytocin binds to its receptor it initiates a cascade of intracellular events that culminate in a range of cellular responses including an increase in neuronal firing, neurotransmitter release, smooth muscle contraction and protein phosphorylation. In rats, peripheral expression of oxytocin receptors is concentrated (but not exclusively) in the male and female reproductive tract and in myoepithelial cells in mammary tissue [4,55]. In addition, oxytocin receptors are also abundantly expressed throughout the CNS and often exist in the same regions containing oxytocin fibers. In addition to their expression in the SON and PVN, oxytocin receptors are also found in the regions of the cortex, hippocampus, limbic system, basal ganglia, medial preoptic area (MPOA), olfactory bulbs, amygdala, and the brain stem [56,57]. There is widespread distribution of oxytocin receptors in the thoracic and lumbosacral segments of the spinal cord, with the dorsal horn, dorsal gray commissure, intermediolateral cell column all possessing oxytocin receptors [58]. However, some brain areas show a distinct mismatch between oxytocin fiber distribution and oxytocin receptor expression, such as seen in the amygdala and olfactory bulbs where there is a significantly greater proportion of oxytocin receptors compared to oxytocin fibers that innervate these nuclei [59–61]. Such an anatomical mismatch gives rise to the possibility that centrally released oxytocin can diffuse to distant sites within the brain to exert its effects. Therefore, oxytocin in the brain is described as a neuromodulator and appears to have broad permissive actions.

Dopamine and Oxytocin Interactions

Stimulation of central dopamine and oxytocin pathways are known to have similar effects on certain social and affiliative behaviors such as sexual behavior and pair bonding [62,63]. In addition to producing similar prosocial behavioral responses, anatomical, and immunocytochemical studies have revealed that the receptor binding sites and neuronal fibers of these two neuroregulators exist in the same CNS regions, often in close apposition to each other [58,64–69]. Furthermore, we have recently shown

that hypothalamic oxytocin cells express dopamine receptors [70] suggesting direct regulation and in the context of sexual behavior, both dopamine and oxytocin activity are known to increase in the same brain region of male rats [71,72]. These observations have led some researchers to believe that central dopamine and oxytocin systems interact with each other to regulate socio-affiliative behavior. As associated behavioral disorders and more profound social deficits are often seen in patients with psychiatric disorders such as autism and depression, it seems logical to assume that disruptions to the integration between dopamine and oxytocin pathways may partly underlie social impairments found in these patients.

Based on this evidence above we will now aim to describe a basic framework of “interaction” between dopamine and oxytocin pathways in a socio-sexual context based on rodent studies, for referring to later in the review (see Figure 2). We provide a general but not exhaustive summary of brain nuclei known to regulate two well-understood social behavior contexts (sexual behavior and pair bonding) and the existence of an overlap of dopamine/oxytocin receptors and projections in these regions. More in-depth neuropharmacological evidence relating to each particular behavioral paradigm will be discussed later in each subsection.

Framework of Proposed Interactions between Dopamine and Oxytocin

Common central brain regions believed to be involved in mediating socio-sexual behaviors include the MPOA, SON, PVN, amygdala, NA, and the VTA. The hypothalamus and limbic system appear to be crucial components for the execution of socio-affiliative behaviors in rodents and for mediating reward pathways as a consequence of social interaction [70,73–76].

Sexual Behavior

The MPOA, SON, and PVN are understood to have roles in regulating penile erection and copulation in male rodents [70,77,78]. These oxytocin-rich nuclei are innervated by dopaminergic fibers from the incertohypothalamic system (located in the zona incerta) (see Figure 2) [64,65] and are known to express dopamine D2-like receptors [70] suggesting a direct regulation of hypothalamic oxytocin by dopamine. In addition to local dendritic oxytocin release from magnocellular neurons [50] the PVN exerts its widespread effects via oxytocin release in other key brain regions notably the hippocampus, amygdala, VTA, and spinal cord to mediate sexual behavior components [73,74].

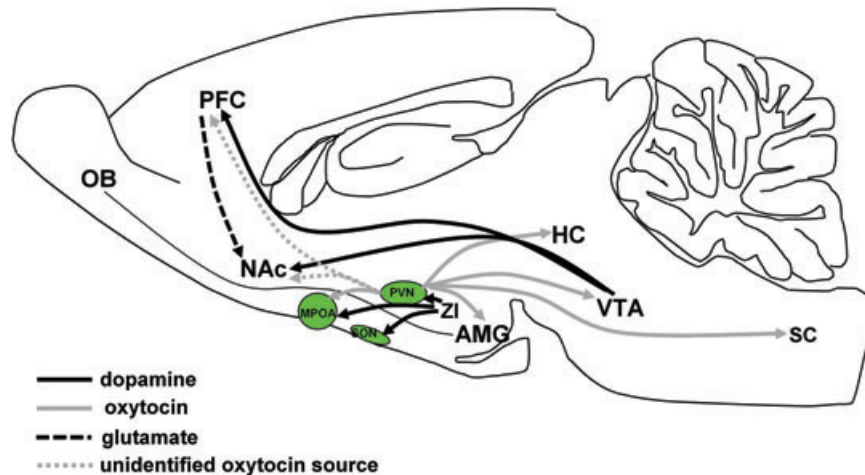


Figure 2 Major dopamine pathways and their relationship to oxytocin neuron populations. Basic framework of proposed interactions between dopamine and oxytocin in the rodent social brain. Sagittal view of a rat brain illustrating potential neural pathways involving dopamine and oxytocin during sociosexual behavior (proposed pathways underlying pair bonding were taken from prairie vole literature as rats do not form pair bonds). Sociosexual behavior is governed by oxytocin release from the hypothalamic nuclei, namely, MPOA, SON, and PVN which receive dopaminergic innervation originating in the ZI. The hypothalamus exerts its pro-social effects using oxytocin via (1) magnocellular oxytocin dendritic release which in turn diffuses throughout the hypothalamus and to other sites and (2) via PVN extra-hypothalamic oxytocin projections to the hippocampus, amygdala, VTA, and spinal cord where they have a role in sexual behavior,

reward and pair bonding. During mating oxytocin release in the AMG, HC, and VTA facilitates social learning and memory and stimulates mesolimbic dopaminergic reward pathways projecting to the NA and PFC. Mating encourages pair bonding possibly via oxytocin release (likely to be supplied by the PVN) in the PFC and the NA, (or in the case of males, vasopressin release in the ventral pallidum, not shown). PFC dopamine levels may increase upon oxytocinergic stimulation leading to further dopamine release in the NA via glutamatergic projections. Concurrently, NA dopamine may also be directly activated by oxytocin to modulate pair bonding. NA, nucleus accumbens; ZI, zona incerta; MPOA, medial preoptic nucleus; PVN, paraventricular nucleus; SON, supraoptic nucleus; AMG, amygdala; VTA, ventral tegmental area; HC, hippocampus; PFC, prefrontal cortex; OB, olfactory bulbs; SC, spinal cord.

The hippocampus and amygdala, which are important in processing social emotions and memory comprise part of the limbic system which also includes the NA, a key site involved in arousal, motivation and reward. The VTA, hippocampus, and amygdala all receive oxytocin input from the PVN [79–81], contain oxytocin receptor mRNA [82,83] and are highly responsive to the pro-erectile effects of oxytocin [73,74]. Furthermore, recent immunocytochemical studies revealed that oxytocin fibers originating in the PVN, lie in close apposition to mesolimbic dopamine cell bodies in the VTA that terminate in the NA [73], thus providing some neuroanatomical basis of a potential paraventricular oxytocin input to mesolimbic dopamine fibers. In addition to the NA [84], the VTA also supplies dopaminergic fibers to the hippocampus and amygdala [85] suggesting that dopamine–oxytocin interactions within these nuclei may also have a bi-directional role.

Taken together this provides some preclinical evidence for an oxytocin–dopamine (and/or dopamine–oxytocin) circuit operating between the PVN, VTA, hippocampus, and amygdala during penile erection. In summary, it has been proposed that during sexual arousal, stimulation of the mesolimbic dopamine system via oxytocin (released

in the VTA, hippocampus, and amygdala) activates in turn incertohypothalamic dopamine fibers innervating the MPOA, SON, and PVN of the hypothalamus. From here oxytocin is believed to act within the hypothalamus, in limbic brain regions and the spinal cord, culminating in the activation of mesolimbic dopamine reward pathways and expression of penile erection (see Figures 2 and 3 for summary).

Pair Bonding

Most studies examining the neural correlates of pair bonding use prairie voles as unlike rats, they form long-lasting bonds after mating and can display high social functioning in certain behavioral paradigms [76,86]. Similar to sexual behavior, the limbic system has a highly integrated role in social attachment behaviors such as pair bonding [66] via its dense projections to the prefrontal cortex, a region known to mediate complex cognitive behaviors. The prefrontal cortex and NA receive dense dopaminergic input largely from the VTA and an oxytocinergic innervation, the source of which is not known, although it is likely to originate in the PVN. In addition, both dopamine and oxytocin receptors are known to be

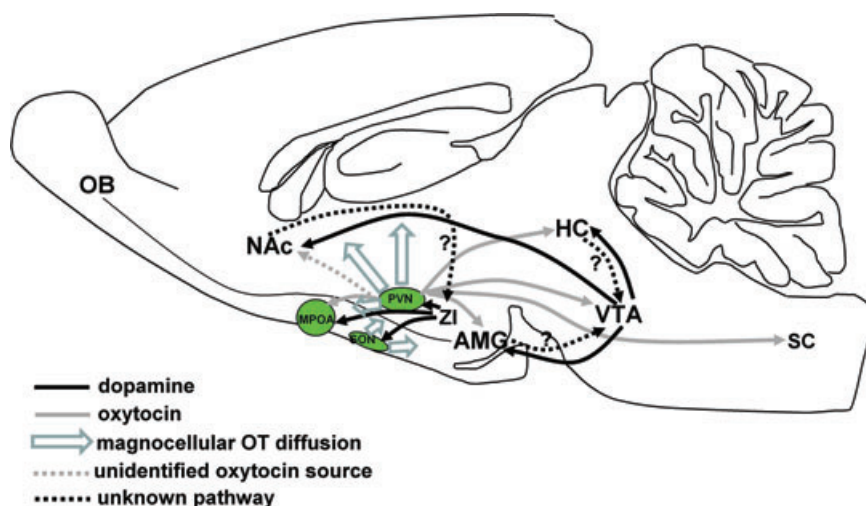


Figure 3 Dopamine and oxytocin interactions during penile erection. Sagittal view of a rat brain illustrating proposed interactions between dopamine and oxytocin in the rodent brain during penile erection. During sexual arousal oxytocin acts in the AMG, HC, and VTA via parvocellular oxytocin projections and magnocellular oxytocin diffusion to stimulate the mesolimbic dopamine pathways originating in the VTA and projecting to the NA, which mediate sexual motivation and reward. Concurrently, mesolimbic dopamine activates (via an unknown pathway) the incertohypothalamic dopamine system to stimulate oxytocinergic neurons in the

PVN which then project to the SC and facilitate penile erection. The role of oxytocin action in the MPOA and SON during penile erection remains unknown, however, they may be involved in mediating those sexual events occurring after erection such as pelvic thrusting and/or ejaculation. OT, oxytocin; NA, nucleus accumbens; ZI, zona incerta; MPOA, medial preoptic nucleus; PVN, paraventricular nucleus; SON, supraoptic nucleus; AMG, amygdala; VTA, ventral tegmental area; HC, hippocampus; OB, olfactory bulb; SC, spinal cord.

abundantly expressed in the prefrontal cortex and the NA [66,87,88]; however, the phenotype of these neurons has yet to be identified. Thus, it may be that, in addition to mesocortical and mesolimbic stimulation during pair bonding, dopamine and oxytocin (via hypothalamic input) may also interact in the prefrontal cortex to modulate dopamine activity in the NA (behavioral neuropharmacological studies are required to confirm this). Concurrently, there may also be coactivation of dopamine and oxytocin in the NA as revealed in a recent study where stimulation of D2-like receptors and oxytocin receptors in the NA facilitated pair bond formation in female prairie voles [89]. Thus, the prefrontal cortex and NA may serve as other potential integrative sites for dopamine and oxytocin pathways underlying natural reward circuits and social attachment behaviors governing for example, maternal, and pair bonding (see Figure 2 for summary).

Sexual Behavior Dysfunction

Male Sexual Behavior

The Diagnostic and Statistical Manual of Mental Disorders (American Psychiatric Association) (DSM-IV) classification for men with sexual dysfunction disorder is the inability to achieve or maintain penile erection until completion of sexual activity. Erectile dysfunction can be sep-

arated into two main categories where the causes are either psychologic (where underlying emotional or mental health processes such as depression and anxiety affect the ability to achieve erection) or organic (where there is a central and/or peripheral disorder in the erectile pathway) in origin. Psychologic, organic causes and a mixture of both comprise approximately 12%, 68%, and 20% of those reporting erectile dysfunction [90]. In our opinion, the most convincing evidence to suggest a central dopamine/oxytocin link in CNS function has originated from preclinical studies investigating male sexual function. The aim of such studies has been to explore a dopamine/oxytocin basis in penile erection in healthy rodents in the hope of better understanding the erectile process in humans which in turn may aid in the development of potential therapies for treating erectile dysfunction. A model of erectile dysfunction has been identified in rats [91]; however to our knowledge, there are no reports in the literature examining or manipulating dopamine/oxytocin pathways in this model. So for this section of the review we will mainly focus on (unless stated otherwise) studies using healthy rodents. In addition, whilst there has been a recent suggestion that dopamine and oxytocin may partly mediate the ejaculatory component of sexual behavior in rats [92], it is the serotonergic system that is generally believed to influence central oxytocin neurotransmission at ejaculation

[93]. Thus, this section of the review will focus on penile erection in rodents and potential therapeutic implications for humans with psychogenic erectile dysfunction.

Mechanisms of Penile Erection

Whilst peripheral processes control contractant and relaxant capacities of corpus cavernosum smooth muscle, penile erection, a spinal reflex, is centrally mediated and involves both spinal and supraspinal pathways [94]. It is a complex response influenced by neural, steroidal, hormonal and vascular inputs [95–97]. Upon appropriate stimulation (e.g., tactile, visual, and auditory) supraspinal (those originating in the PVN) and peripheral pathways converge on spinal pro-erectile centers to activate autonomic fibers running in cavernosal nerves, which provide the neuronal efferents to the penis. This integrated and coordinated input into the spinal cord culminates in engorgement of the penis with blood and penile rigidity (enhanced by contraction of the perineal striated muscles).

In healthy human males, circulating oxytocin levels in the blood are known to rise during sexual arousal, penile erection and ejaculation [98,99]. Similarly in male rats, peripheral and either intracerebroventricular (i.c.v.) or microinjection in various brain regions (including the amygdala, hippocampus, VTA, PVN, and lumbar spinal cord) of oxytocin facilitates penile erection and copulation [10,73,74,100,101]. Both rodent and human studies suggest that oxytocin acts as an important mediator for both appetitive and consummatory phases in male sexual behavior. In addition to oxytocin, dopamine is also known to partly comprise those excitation pathways governing the expression of sexual behavior. Dopaminergic agonists have been shown to exert erectogenic effects in clinical studies [102]. Similarly, when administered locally into the MPOA or PVN in the male rat, dopamine receptor agonists exert a facilitatory effect on almost all aspects of male sexual behavior, particularly penile erection [77,103–105]. Such findings have led researchers to believe that central dopaminergic and oxytocinergic pathways may interact with each other to mediate erectile function; however, the nature of interactions and brain circuits employed by dopamine/oxytocin are only beginning to be probed.

Morphological and Electrophysiological Evidence

As mentioned previously in the rat, the oxytocin-rich MPOA, SON, and PVN of the hypothalamus receive dopaminergic input [64,65] and are believed to be critical integrative sites for male sexual behavior [77,78,106] (see

Figures 2 and 3). We have recently shown that oxytocin cells in the MPOA, SON, and PVN possess dopamine D2, D3, and D4 receptors [70] which suggest that dopamine may be able to directly influence hypothalamic oxytocin neurotransmission via D2-like receptors. The ability of dopamine to positively influence hypothalamic oxytocin release was first demonstrated in early *in vitro* studies [107]. Since then, there have been several *in vivo* studies demonstrating a marked increase in oxytocin concentration in the blood, hippocampus and PVN in response to dopaminergic stimulation [41,108–111]. Electrophysiology has also shown that specific targeting of D1, and D3 receptors can influence the depolarization of oxytocin cells in the hypothalamus [112,113] which further confirms the dopamine receptor(s) involved in central oxytocin release.

Behavioral Pharmacological Evidence

Behavioral pharmacological studies in rats have been extremely informative and insightful by revealing a strong link between central dopamine and oxytocin neurotransmission in the context of sexual behavior. The relationship between dopamine and oxytocin neurons in the PVN during penile erection was substantiated by the attenuation of apomorphine-induced penile erection after bilateral lesioning of the PVN [114] thus depleting extra-hypothalamic oxytocin stores [115]. Stimulation of penile erection by apomorphine or the selective D4 receptor agonist, PD 168077 is also prevented by i.c.v., but not intra-PVN delivery of an oxytocin receptor antagonist [104,105,116] suggesting that oxytocin receptors located outwith the PVN are involved in mediating apomorphine-induced penile erection. Dopamine agonist-induced penile erection can be inhibited by oxytocin receptor blockade [114]; however, penile erection elicited by oxytocin is not inhibited by dopamine receptor blockade [117] suggesting that dopamine may lie upstream to oxytocin pathways. More recently, this finding was contradicted after Martino et al. [101] revealed the pro-erectile effect of oxytocin was inhibited by the dopamine receptor antagonist, clozapine, which suggests that oxytocin may be able to modulate central dopamine neurotransmission in regulating erectile function. To add to the neurochemical complexities subserving penile erection, we have recently shown that dopamine may differentially stimulate oxytocin subpopulations to produce erectile events. This dissociation appears to be dependent on whether intromission (physiological marker of penile erection where male rat positions himself behind and mounts receptive female leading to a train of pelvic thrusts, termed *in copula* penile erection) is achieved. In our study, we found naive rats

administered with the D2/D3 agonist, Quinelorane but not the D4 agonist, PD168077 (i.c.v.) displayed multiple erectile episodes associated with the activation of parvocellular oxytocin neurons but had no apparent effect on magnocellular oxytocin cells. In a separate study, we aimed to block endogenous dopamine release in sexually experienced males allowed full access to receptive females by i.c.v. injection of a D2, D3, or D4 dopamine antagonist. We found that blockade of central D4 receptors was the most effective at inhibiting intromission and this was correlated with attenuation of magnocellular oxytocin neuron activation [70] suggesting that *in copula* erection in rats may be D4 receptor-mediated.

It is not clear as to whether during penile erection dopamine increases oxytocin levels in the PVN (via magnocellular cells) and/or at sites outwith the PVN (via parvocellular cells) such as in the spinal cord where pro-erectile centers exist [58] or the hippocampus [108]. There is increasing evidence to suggest that parvocellular oxytocin neurons are part of the neural network controlling penile erection [108,118,119]. Apomorphine-induced penile erection involves, at least in part, release of oxytocin at extrahypothalamic areas via these parvocellular fibers; however, intrahypothalamic oxytocin release and action cannot be ruled out. In agreement with this we have recently demonstrated in anesthetized rats, that blockade of oxytocin receptors in the lumbosacral spinal cord significantly reduces the facilitatory effect of intravenous (i.v.) injection of apomorphine on intracavernous pressure rises (ICP, physiological index of penile erection) leading us to believe that dopamine may activate a paraventriculospinal pathway in the generation of erection [70]. Contrary to these findings, we were unable to show that apomorphine (i.v.) markedly affected spinal oxytocin release in the lumbosacral spinal cord (unpublished observations), which leads us to conclude that spinal oxytocin may have only a minor modulatory role in apomorphine-induced penile erection and there is involvement of other neuromediators. Indeed, central oxytocin receptors are highly homologous to another key neuropeptide, vasopressin [4]. Such similarities in receptor structure will undoubtedly influence ligand receptor recognition/activation. In the spinal cord for example, conflicting data has revealed that oxytocin may exert its effects via vasopressin [120] or oxytocin receptor activation [121], suggesting a role for central vasopressin in regulating male sexual behavior cannot be ruled out.

Potential Brain Circuits

Some recently published animal data has implicated involvement of other brain nuclei in the complex communication network which uses central dopamine

and oxytocin systems to mediate rodent sexual behavior (see Figure 3). Oxytocin stimulation of the ventral subiculum in the hippocampus, medial amygdale or VTA, and apomorphine microinjection in the PVN all induce penile erection preceded with a marked increase in mesolimbic dopamine activity suggesting an oxytocin–dopamine driven pathway. Both responses were inhibited after blockade of central oxytocin receptors and hypothalamic and mesolimbic dopamine receptors [73,74,110,122]. Thus, such findings suggest that not only dopamine–oxytocin but oxytocin–dopamine pathways may subservise penile erection but also implicates a role for dopamine and oxytocin in the activation of pathways governing sexual motivation and sexual reward. Taken together, these findings suggest dopamine/oxytocin interactions underpinning erectile function involve dopamine and oxytocin-containing nuclei within and outwith the PVN that form a much larger and highly integrated network subserving appetitive and consummatory components of sexual behavior (see Figure 3 for summary).

So referring to our framework for interaction, a substantial amount of evidence of dopamine and oxytocin projections, targets and receptor localization, and sophisticated pharmacological studies, allow us to conclude that dopamine–oxytocin (and oxytocin–dopamine) interaction is important for the normal display of penile erection.

Treatment of Erectile Dysfunction

In humans erectile dysfunction is characterized by the inability to achieve and maintain penile erection sufficient to engage in sexual intimacy and affects around 2 million men in the United Kingdom. Despite other contributing factors such as age, prevailing underlying vascular diseases and psychogenic factors, the causes of erectile dysfunction are in most cases due to a disruption in either or both peripheral and central pathways controlling penile erection. Whilst the use of nonpharmacologic therapies such as vacuum erection devices, penile implants and psychotherapy have reported some success in patients unable to achieve erection [123]; pharmacotherapy targeting peripheral (and in some cases central) sites currently remains first-line treatment. Currently, the intracellular messenger nitric oxide (NO) has been receiving some attention due to its ability to act on postganglionic parasympathetic fibers innervating penile smooth muscle and so aid in initiating and maintaining erection. Indeed, pharmacological agents acting to replenish NO have shown some success in preclinical and clinical studies of erectile dysfunction; however, research is still at an early stage [124]. The dopamine agonist, apomorphine

did show some clinical success due to its ability to activate pro-erectile sites in the CNS and induce erectile responses [125,126]. However, there was limited patient tolerance due to adverse side effects [127]; some too severe to provide any long-term treatment. Intranasal delivery of apomorphine may be a potentially effective alternate route of administration since it facilitates preferential delivery to the brain [128], was shown to be a powerful sexual stimulant in male rats and was also well tolerated by patients without adversely affecting cardiovascular function or inducing nausea [129,130]. However, the effectiveness of intranasal apomorphine as a powerful erectogenic without substantial adverse side effects warrants further investigation.

Oxytocin has become a key clinical target due to its potent erectogenic effects in preclinical studies [9,101] and seemingly lack of severe side effects when given in low doses, e.g., to pregnant patients to induce labour [131]. However, to date there are no oxytocin agonists clinically available for the treatment of erectile dysfunction. According to the literature, at present, there do not appear to be any clinical studies exploring the effects of oxytocics in patients with erectile dysfunction and this is presumably due to the greater clinical outcome of other pharmacotherapies such as those targeting the melanocortin system. Bremelanotide, a melanocortin agonist, is one of the first centrally acting agents shown to have greater clinical efficacy and patient satisfaction than currently available treatments [132,133]. Interestingly, bremelanotide is likely to act via oxytocin as oxytocin neurons express melanocortin receptors and the endogenous melanocortin, alpha melanocyte stimulating hormone activates oxytocin neurons and facilitates sexual behavior in rats [106]. Phosphodiesterase-5 inhibitors (PDE-5) (such as Viagra) are currently the safest treatment for patients with erectile dysfunction with fewer contraindications [102,126,134] and provide overall greater patient satisfaction [135]. However, they act peripherally and are effective in maintaining penile erection in patients capable of initiating sexual arousal; however, they are not able to induce penile erection. Thus, those patients where psychogenic factors are a contributing factor would benefit from treatments aimed at targeting the CNS. Patients suffering psychological disorders such as depression and anxiety or neurologic conditions such as Parkinson's disease, where there is profound disruption to central dopamine pathways (and presumably oxytocin neurotransmission), frequently report bouts of erectile dysfunction [136,137]. Thus, development of specific dopaminergic ligands (specific for D2-like receptors) or oxytocics which would act to redress central dopamine and oxytocin levels in the CNS may aid in the treatment of patients reporting decreased li-

bido. However, administering dopaminergic agents aimed at selectively targeting for example, central D2-like receptors does not appear to be more advantageous since the unwanted peripheral side effects still persist. The lack of biological receptor specificity to delineate central and peripheral (located in the area postrema in the brain) receptors remains a huge hurdle for pharmacotherapeutic development.

Development of novel pharmacotherapies targeting the central oxytocin system is at a preliminary stage [138]; however, issues with regard to accessing CNS sites has proved problematic since oxytocin does not readily pass the blood-brain barrier. Delivery into the CNS via intranasal administration of other neuropeptides such as vasopressin has been reported in humans [139], furthermore, intranasal delivery of oxytocin was shown to exert antidepressant effects in humans [140], suggesting this could be one route of administration whereby oxytocin can act centrally. It is too premature to speculate on the potential therapeutic value of oxytocinergic stimulants; however, activation of the central oxytocin system may offer some promise and potential benefits over viagra, especially in those patients who experience difficulties in arousal/initiating penile erection. Thus, there is a need for more explorative research to aid in the development of safe and effective treatment for erectile dysfunction.

Female Sexual Behavior

The DMS-IV classification of female sexual dysfunction is defined as a "disturbance in sexual desire and in the psychophysiological changes that characterize the sexual response cycle and cause marked distress and interpersonal difficulty." In women, there are four main categories, which are used to define female sexual disorders: sexual desire, sexual arousal, sexual orgasmic, and sexual pain disorders [141]. However, the criteria used to diagnose female sexual dysfunction remains a contentious subject, as unlike men, there is not a quantifiable physiological index that can be measured in women. In addition, the female sexual response is highly sensitive to emotional and psychosocial states, both of which can influence sexual desire, arousal, and gratification. Thus, attempting to understand the neurobiology underlying the different facets of female sexual function using rodent models remains very difficult, however, they are now beginning to be explored [142]. Unlike penile erection, identifying a potential link between central dopamine and oxytocin pathways during female sexual behavior has yet to be investigated in preclinical studies and to our knowledge animals model of female sexual dysfunction have yet to be established. Sexually receptive female rats display proceptive behaviors which are comprised of "ear wiggling,"

“hopping,” and “darting” (analogous to sexual arousal in women) to generate male attention and facilitate sexual activity. Female rats then display receptive behaviors, which include lordosis, a supraspinal reflex (believed to be analogous to orgasm in women), which is expressed in response to tactile, e.g., mounting male rat. Here the female displays a characteristic posture by arching the back moving the tail to allow penetration by the male rat) [77,143]. The involvement of dopamine and activation of central brain regions containing oxytocin cells during sexual behavior paradigms in female rats [142] suggests similar neurochemical processes involved in male sexual behavior may also be operative in females. Similar to those key regions comprising the basic framework for sexual behavior in males (see Figure 3), the MPOA, PVN, and amygdala are activated upon sexual stimulation; however, the ventromedial hypothalamus, bed nucleus of stria terminalis and lateral septum have also been implicated in the expression of female sexual behavior (not included in basic framework in Figure 3) [144–146].

Clinical studies have reported that plasma oxytocin levels in healthy women are markedly elevated during sexual arousal, orgasm, and remain high during the refractory period immediately after orgasm suggesting a role for oxytocin in female appetitive, consummative and post-coital behavior [98,147]. In female rats, sexual behavior culminating in lordosis coincides with activation of oxytocin cells in the ventromedial hypothalamus believed to be supplied by neurons in the PVN [144]. In gonadal steroid primed females, microinjection of oxytocin into the MPOA, VMH, or lateral ventricle facilitated the expression of female sexual behavior which was subsequently inhibited after central administration of an oxytocin antagonist [148–150]. As in humans, understanding the neural basis of female sexual behavior in rodents is further complicated by the basal levels of circulating oestrogen and progesterone (depending upon stage of oestrus cycle) and may help explain some discrepancies in the literature regarding the prosexual effects of oxytocin in females [150].

The role of dopamine in female sexual function is more ambiguous than in males. Some findings have shown dopamine to participate in the activation of those pathways controlling sexual motivation, sexual arousal and sexual reward in females [77,142]. However, its involvement in female sexual reflexes is less well defined. Early studies suggested that dopamine may have an inhibitory role in females as inferred by studies using dopamine agonists such as quinlorane and analyzing their effect on lordosis. The authors found that stimulation of central dopaminergic systems (although which pathways is unknown) reduced the expression and duration of lordosis [151,152] whilst others reported the opposite

[153,154]. Lesioning of dopamine-rich regions such as the MPOA and microinjection of dopamine antagonists into the MPOA markedly disrupts appetitive behaviors in female rats. Selective blockade of MPOA D1 receptors in particular attenuates vaginocervical stimulation-induced neuronal activation [155,156]. These findings suggests dopamine action in the MPOA is important in the sexually active female rat. Furthermore, microdialysis has revealed an increase in dopamine levels in the NA in female hamsters and rats in the presence of a sexual stimulus (exposure to unaccessible male) and during mating [157,158] which is similar to those observations in males. As with oxytocin, dopamine's role in female sexual behavior is dependent on the hormonal treatments administered to female rats, which can clearly influence the effect of dopamine agonists on sexual behavior. Lordotic responses in nonreceptive and receptive rats appear to differ after dopaminergic stimulation [153,159]. Thus, our understanding of dopamine's involvement in component behaviors of female sexual activity is more complex and requires more thorough investigation.

To our knowledge, there are no data available examining an interaction between dopamine and oxytocin during female behavior studies in rodents. It is tempting to suggest that similar cross-talk between dopaminergic and oxytocinergic processes may subservise components of female sexual behavior as seen in males since disruption to the MPOA, known to be highly responsive to the prosexual effects of both dopamine and oxytocin in male rats, attenuates expression of sexual behavior in females. It may be that as indicated in males, the MPOA may form part of larger circuit involving oxytocin/dopamine interactions extending to key nuclei in the limbic system such as the VTA and NA to mediate the expression and reinforcement properties of lordosis. Therefore, there is not enough evidence to say yet that female proceptive or receptive behavior relies on a dopamine–oxytocin interaction according to our framework, although some elements clearly depend upon one or other factor. More explorative studies have to be carried out to confirm the role of oxytocin in dopamine-mediated processes governing female sexual behavior.

Female Sexual Dysfunction and Treatment

There are currently no therapeutic interventions available for women suffering sexual desire/arousal disorders. Similar to males, some clinical success has been reported with the CNS-acting agent, bremelanocortin where females reported enhanced sexual desire, arousal and significantly greater satisfaction during intercourse [160,161]. This suggests the melanocortin system may be a promising candidate and of potential therapeutic value.

Augmentation of the serotonergic system appears to be another area of sexual pharmacology that is of interest. The serotonin agonist, flibanserin (targets 5-HT_{1A} and 2A receptors) has reached clinical trials and acts to dampen the serotonergic system and redress dopaminergic and noradrenergic brain levels in a dual manner by decreasing serotonin levels and inhibiting cortical serotonergic neurotransmission [142 and references therein]. Because preclinical studies are at such a preliminary stage investigating the role of dopamine and oxytocin pathways in female sexual behavior, it is difficult to draw conclusions regarding any potential treatment options targeting these two systems. However, it could be postulated that since oxytocin release in women has been positively correlated to romantically linked affiliation cues, sexual arousal, and orgasm [98,162], similar parallels may be drawn in females from clinical trials in males, because similar dopamine and oxytocin systems appear to underlie female and male sexual components; however, currently it is too early to say.

Social and Parental Behavior and Related Disorders

Socially related behaviors other than sexual behavior include social bonding and maternal behavior, each of which comprise recognition, memory, seeking close proximity to conspecifics, and other behavioral components. There are some well-described functions for both oxytocin and dopamine in the control of these social parameters. The fundamental evidence for oxytocin's role is that oxytocin- or oxytocin receptor-null mice exhibit deficits in social recognition and social memory [25,163–165] indicating the necessity of oxytocin in facilitating interaction between individuals. Oxytocin in the brain also has antistress and antianxiety roles [166], further showing its pleiotropic nature in handling social situations. Similarly, disruption of dopamine signalling in transgenic mice also leads to social abnormalities [167,168], e.g., dopamine transporter null mice, that have increased extracellular dopamine and decreased dopamine receptor expression, exhibit increased social reactivity. We will analyze the reciprocal role that oxytocin and dopamine play in such behaviors in rodents and humans and then discuss whether disorders of social behavior, such as autism, can be explained by oxytocin and/or dopamine dysfunction.

Social Bonding

Social bonding includes elements such as time spent together in close proximity, choice of the known individ-

ual over another unknown individual (social preference), and social recognition. Social recognition is experimentally tested in the lab by measuring the time spent investigating a known individual compared to an unknown one and relies on olfactory and visual cues. Olfactory learning is a major component of recognition, involving noradrenaline-mediated disinhibition of mitral cells [169]. However, olfactory and visual cues also both use oxytocin [170,171]. Unlike wild-type mice oxytocin null mice cannot remember conspecific mice that they have recently been exposed to. Oxytocin receptor is expressed in the olfactory bulb, and although the PVN is considered a major source of oxytocin mediating social behaviors, oxytocinergic fibers have not been observed to appose either mitral or glomerular cells so the origin of oxytocin must be from other release sites, e.g., magnocellular dendrites. Other centers for oxytocin action in social recognition include the lateral septum, medial amygdala and MPOA, which also express oxytocin receptor. Since oxytocin can act via noradrenaline or other classical neurotransmitters (glutamate, GABA) in some areas, it is thought to modulate olfactory learning rather than directly stimulate the olfactory substrate [26], so as dopamine also plays a role in the olfactory bulb in social recognition oxytocin may also modulate dopamine release in order to mediate its effects.

Dopamine modulates odour detection and discrimination, which is part of olfactory memory formation [170,172,173], although others argue that dopamine (D₂) plays a more prominent role in consolidation of memory rather than recognition *per se* [174]. So both oxytocin and dopamine play a role within the olfactory bulb in social recognition/memory. The Bruce Effect is an example of the consequences of lack of social recognition; in mated female mice, implantation failure occurs in response to pheromones from a stranger male; both oxytocin and dopamine action in the olfactory bulb are implicated in this phenomenon [175]. However, due to lack of evidence for their interaction within the olfactory bulb, attempts to fit our proposed interaction framework to this behavior is difficult. The precise phenotypes of neurones expressing dopamine or oxytocin receptors are not reported and the source of dopamine or oxytocin in the olfactory bulbs is unclear, while pharmacological approaches have not yet targeted the olfactory bulb selectively. The evidence suggests that there is a local synergistic, if not interactive, effect of oxytocin and dopamine within the olfactory bulbs, but there may also be interaction upstream, in other dopamine or oxytocin source or target sites.

Associated with social recognition, bonding develops under certain circumstances. Animals can develop a close association that is measurable by the social preference

test. Bonding behavior between adults has been closely studied over the past decade using prairie voles as a model species since they develop a well-defined partner preference. When male and female prairie voles spend time together partner preference increases. However, after copulation partner preference is strongly elicited, and the paired voles attain a monogamous-like affiliation. There is even an associated induction of parental behavior in male prairie voles which will be discussed later. This remarkable postcopulatory bonding between adults is attributed in part to both dopamine and oxytocin in both females and males [176], and the NA has been identified as the main forebrain center for the effect. As discussed below, this phenomenon represents another example of dopamine–oxytocin interaction in a behavioral context.

Importantly, the partner preference induction is dependent upon oxytocin receptor distribution in the brain. The NA and the caudate putamen have high densities of oxytocin receptors in animals where partner preference is induced but not in other species, such as the montane or meadow vole or mouse [176]. Oxytocin release increases in the NA in female voles during pairing [68], although the source of oxytocin is unclear. Increased social contact is inducible by intra-cerebral oxytocin infusion and can be induced not only in voles but also in rats and squirrel monkeys; and partner preference is blocked by oxytocin antagonist infusion into the NA. After mating oxytocin release also increases in the PVN and extrahypothalamic regions such as the amygdala in male and female rats [72] (and see above), so conceivably these regions could also play a role in partner bonding. Interestingly mating-induced oxytocin release in rats has been shown to underlie postcopulation anxiolysis so if also true for other rodents this may be a component of the bonding behavior. Whether this occurs in the prairie vole model is unknown and pair bonding in rats is not typically induced by copulation.

The NA is also an important center for dopamine action during pair bond formation in prairie voles [176]. Mating increases dopamine turnover in the NA in males and females- NA neurones are known to express dopamine receptors and dopamine acts via D2 receptors to facilitate partner preference. The phenotype of dopamine target neurones is not described but dopamine presumably acts by regulating the predominant NA GABA neurones, although some evidence indicates that it also modulates incoming glutamatergic afferents [177]. As for oxytocin, dopamine D2 agonists facilitate and antagonists attenuate partner preference behavior.

There is a reasonable body of evidence revealing an oxytocin–dopamine interaction in partner preference. The NA is rich in both oxytocin and dopamine recep-

tors; if coexpressed pre- or postsynaptically it would provide a clear basis for coregulation of the same target neurones, but this is unknown. As well as acting in the NA it appears that mating-dependent release of oxytocin activates a mesolimbic (VTA) dopamine circuit and induces dopamine release in the NA, indicating upstream regulation of a dopamine circuit. Therefore, in association with copulation oxytocin and dopamine link the state of sexual arousal with that of bonding [74]. However, it has not been shown that oxytocin release increases in the VTA where it might stimulate dopaminergic projections at copulation or with pair bonding, although, neuroanatomical evidence has proved indicative (Melis et al., 2007). On the other hand, recent reports propose dopamine acts via oxytocin since concurrent reciprocal activation of dopamine and oxytocin receptors in the NA occurs [89,176]. Pharmacological studies are needed to tease out the precise roles of oxytocin and dopamine in the NA and VTA. One might also ask whether dopamine action on PVN (or SON) oxytocin neurones is necessary for induction of partner preference. Evidence in the prairie vole is lacking but we would speculate that it is part of the complex cascade leading to arousal and social interaction that precedes copulation and bonding.

It should, however, be noted that another hypothalamic peptide related to oxytocin, vasopressin, has also important interactions with dopamine in the NA in social recognition and the formation of pair bonding [176]. There is substantial evidence of prominent vasopressin V1a receptor expression in the NA and ventral pallidum that facilitates the formation of pair bonds and this dependent upon a specific sequence in the 5' flanking region of the gene that determines its brain expression pattern. Vasopressin neurones lie adjacent to oxytocin neurones in the PVN and SON, and also express D2-like receptors [70], so dopamine may control vasopressin release and action in parallel with oxytocin. Whether vasopressin also controls dopamine neurone activity or release (as oxytocin does, see above) is unclear at present. Interestingly though, viral vector insertion of the prairie vole V1a receptor into the mouse brain induces susceptibility to the formation of pair bonds similar to that seen in prairie voles [178].

So, it appears that robust oxytocin–dopamine and/or dopamine–oxytocin connections are involved in social interactions. In mating it may be that dopamine–oxytocin connections are initially involved in penile erection [70], (see above; Figure 3) but then oxytocin-driven dopamine effects mediate subsequent related behaviors such as bonding and reward. However, it is apparent that their interaction at multiple brain locations matches with our framework, and occurs in a social context with more than one potential outcome: copulation and bonding.

Studies of the roles of oxytocin and dopamine in social behaviors in humans are appearing more now with the application of functional magnetic resonance imaging (fMRI) and gene expression studies on post mortem tissue. For example, it can be observed that high intensity fMRI signals are observed in the VTA and SN upon viewing a loved partner [179], and these correlate with the distribution of human oxytocin receptor expression and an interaction with the dopamine system [180]. Other strategies include analysis of peripheral oxytocin concentration [98,181]. Although these do not necessarily correlate with the release or action of brain oxytocin, some importance is attached to increasing levels that correlate to positive mood and prosocial behaviors and, along with studies investigating the effects of intra-nasal administration of oxytocin, have gained some credibility in recent years [182].

Lack of social recognition and inability to form social bonds are characteristics of a host of psychiatric disturbances, the most profound perhaps being autism. Autism and autism-spectrum-disorders represent a profound disturbance in the ability to form social bonds in humans and are most commonly found in males. Although underlying causes are multiple, and include a variety of potential genetic mechanisms, naturally associations between oxytocin and dopamine in autism have been sought to explain the condition.

Autism

The DSM-IV definition of autism is the inability to socially interact due to a deficit in verbal and nonverbal communication, social awareness and interactions and imaginative play. Autism-spectrum-disorders are complex neurodevelopmental disorders characterized by social withdrawal [183] and no adequate animal models have yet been reported. A role for oxytocin has been implied since plasma oxytocin concentrations (which can be a marker for social behaviors in humans) [98,181] are low in autistic boys [184,185] and oxytocin infusions or intranasal administration improve emotion recognition and facilitate trust analyzed using fMRI studies in humans, particularly revealing the amygdala as a target [186–188]. Additionally there is a proposed link between oxytocin receptor polymorphisms and autism in some families [182,189–191], which might adversely alter expression patterns and densities in a way that contributes to the altered social behavior. However, like bonding, autism is also associated with polymorphisms in the vasopressin receptor 1A gene, particularly in the amygdala [192]. Overall the evidence for oxytocin is contradictory, as discussed in more detail in an excellent review by Hammock and Young (2008) [176]. For example, in one study

autistic children with circulating oxytocin levels in the normal range were the most profoundly asocial, so peripheral hormone levels may not reflect central oxytocin release or action in this condition. In contrast to oxytocin, stronger evidence links dopamine dysfunction with autism-like disorders. Reports are mixed but variations in the dopamine system such as dopamine transporter and D4 receptor genes and activity are implicated. Attention deficit/hyperactivity disorder, one of the range of autism-spectrum-disorders, also appears to exhibit similar dysfunction of dopamine systems [193–197]. Whether there are parallel changes in oxytocin release and dopamine activity in relevant brain regions, such as the NA, VTA or ventral pallidum in people with autism-related disorders is uninvestigated, but a joint role for oxytocin and dopamine seems possible.

Treatments for Autism-Spectrum Disorders

Despite the relatively weak evidence for a role for oxytocin in autism, oxytocin treatment has been reported to successfully reduce some characteristic behaviors in autistic patients [198]. Since it is likely that intranasal oxytocin may pass the blood brain barrier [139], there have been some reports of humans showing improvement in communicating behavior and secure relationship attachment [182,199]. Therefore such an approach can potentially be developed into therapies that abrogate some of the more overt social behavioral deficits in autism. On the other hand, drugs targeting the dopamine system such as risperidone and olanzapine seem relatively effective [200]. It may be that combined therapy targeting oxytocin and dopamine systems together would be synergistic, resulting in more effective therapy.

Maternal behavior also creates a bond between mother and offspring akin to that of the social adult-adult bond so we will now outline the parallels between social and maternal behavior and the relative contributions of oxytocin and dopamine.

Parental Behavior

Oxytocin and dopamine are also key neuromodulators of maternal and paternal behavior. It has been recognized for many decades that oxytocin in the brain plays a key role in maternal behaviors [201], and is mainly facilitatory rather than essential, unlike for other social behaviors as discussed above [163,165,202]. More recently, reports of its role in paternal behaviors are emerging, including in prairie voles where mating not only induces social bonding but also paternal care behaviors in males [203–206]. Similarities between mechanisms in mothers and fathers include enhanced oxytocin expression in the

PVN in paternal compared to nonpaternal males and increased vasopressin V1a receptors in the prefrontal cortex in the male marmoset [207]. However, maternal behavior is much better investigated and understood, especially the role of oxytocin in sheep and rodents, so this section will concentrate on evidence from females.

Initially oxytocin was considered to be important for onset of maternal care perinatally but is now recognized to also maintain the suite of behaviors involved. Thus, central oxytocin facilitates care behaviors (pup licking, grooming and nesting behavior), delivery of nutrition to offspring (particularly the milk provision via the ejection reflex for which oxytocin is essential [163], and arched back nursing [kyphosis]), offspring protection (against predators but also including maternal aggression against conspecifics) and, importantly, bonding [26,201,208,209]. Some of these social-like behaviors are comparable to the adult-adult bonding as discussed above, and involve recognition mechanisms.

Mother-infant recognition may be similar to adult-adult recognition in that oxytocin mediates olfactory memory for offspring. This is very well investigated in sheep, where olfactory memory is a crucial, postbirth link between ewe and lamb and is necessary before any further behaviors, including allowing lamb suckling, are performed [26,170]. Such olfactory memory is part of mother-infant bonding in sheep that occurs at birth and can be induced by vagino-cervical stimulation under appropriate steroid conditions. Bonding between ewe and lamb does not occur in the absence of vagino-cervical stimulation, and even in women it has been reported that after caesarean birth, bonding with the baby takes significantly longer than after vaginal delivery [210]. Bonding combined with olfactory memory of the young initiates the other maternal caring and nurturing behaviors [26]. Initially, birth and vagino-cervical stimulation increase oxytocin release in various brain regions, including the PVN, SON, MPOA, SN, septum and olfactory bulb in both rats and sheep [26,170], which are known to be key regions mediating maternal interaction from a variety of activity and lesioning studies. Mimicking this endogenous release pattern with central administration of oxytocin induces, and oxytocin antagonist inhibits, maternal behaviors in rodents and sheep. However, as for prairie vole bonding, oxytocin receptor distribution is most important in quality of behavior performed. Not only does oxytocin receptor expression increase perinatally and into lactation [201], but 'good' maternal behavior, commonly defined as high licking, grooming and arched back nursing, correlates with wider and higher density of distribution of oxytocin receptor in a rat model [211]. Furthermore, such good behavior can be epi-genetically inherited by daughters, as care received neonatally deter-

mines their brain oxytocin receptor distribution and maternal behavior quality in adulthood [212]. Therefore receptor patterns rather than strictly quantity of neuropeptide release may be particularly important in determining quality across the range of social behaviors. Maternal experience is important too: stress exposure perinatally alters oxytocin receptor expression patterns correlated with poorer behavior [213], and this may also extend to stress emanating from prolonged conditions relating to a difficult (e.g., psychological disturbances) or ill child. The best evidence links oxytocin to maternal behavior in women in a correlative way, e.g., fMRI imaging of the maternal brain while viewing photos of their baby reveals activation of oxytocin target regions (e.g., SN) [214], and increased CSF or plasma oxytocin levels [26,215] are evident compared to women who are not mothers. Further indication of a role for oxytocin in primates comes from central administration of oxytocin to rhesus monkeys which increases maternal behaviors [216], and oxytocin is widely believed to be evolutionarily important in maternal care across a range of species.

Proof of the importance of dopamine activity in mothers' brains was shown in dopamine transporter knockout mice which exhibited impaired maternal behavior [217,218]. Like oxytocin, dopamine release is also elevated in the PVN, SON, MPOA, SN, septum and olfactory bulb in rats and sheep [26,170]. Unlike oxytocin, where receptor distribution is primarily important, levels of dopamine in the NA equate with quality of maternal behavior [219]. Lesioning the VTA, using 6-hydroxydopamine to selectively destroy monoamine cells, also blocks maternal behavior [26] indicating that as a potentially important source of dopamine in mothers' performance. Evidence indicates that extensive interaction between oxytocin and dopamine plays a role.

Oxytocin modifies targets directly and/or presynaptically, at least partly via monoamines, including dopamine, and GABA [26], so could potentially gate dopamine effects. At birth oxytocin receptor increases in many classical dopamine target regions, including NA, olfactory bulb, and prefrontal cortex, increasing the potential for oxytocin regulation of the dopamine release at onset of maternal behavior [26]. However, oxytocin receptor expression evidently decreases in the SN, showing that control of the nigrostriatal dopamine neurones and their emanating pathway(s) is differentially altered from other dopamine sources. Oxytocin released after birth or vagino-cervical stimulation modulates dopamine activity in the NA, VTA and SN, and dopamine reciprocates with action within the PVN [26]. It has been proposed that oxytocin action on dopamine neurone activity in the SN promotes immobility to facilitate offspring suckling, but the coregulation is also associated with behavioral drive

as well as feelings of reward associated with the neonate [208]. How this equates with decreased oxytocin receptor in the SN is unknown, but may indicate a shift in oxytocin control from one mediator to another within the SN. Interestingly, mothers showing better maternal behavior (high licking and grooming) have increased oxytocinergic projections to the VTA and more dopamine release within the NA, powerfully revealing the importance of the oxytocin–dopamine interaction located in the mesolimbic pathway [220]. So there are similarities between maternal bonding and adult bonding in the neurochemical oxytocin–dopamine interaction but corresponding experiments have not all been performed to clearly compare them or to match to our framework. Interestingly, recognition of the mother by the offspring (rat pups) also involves monoamines [221], and others have speculated that oxytocin action in the offspring brain might parallel that of the mother [176].

Disorders of parental behavior are not yet well described, let alone understood physiologically, but have profound consequences on offspring. One topical and emerging field is that of offspring adverse programming arising from parental and/or postnatal stress. Adverse neonatal programming can also be produced by poor parental care, whether or not stress is manifest. Such adverse long term consequences on offspring psychological and physical development arising from parental stress or poor parental care in the neonatal period include susceptibility to anxiety and depression disorders, obesity, and cardiovascular disease [222]. In fact, perinatal stress and level of maternal attachment also has an impact on dopamine systems in the offspring, detrimentally altering its release and activity parameters and reproductive development [223,224].

So, abnormal or inappropriate parental behavior in humans has far-reaching consequences for children and, for offspring, perinatal experience received has a huge impact on their later development. As indicated earlier, there is also a reciprocal effect, where disruptive or socially compromised children inflict stress and long-term consequences on the parents and their behaviors. This is only just being recognized as a problem for families, and will have costs for the health and social services in the future. Understanding of the causes of poor parental behavior is only just emerging, but includes parents receiving poor parental care when they were young and peri-natal stress exposure, setting up a vicious cycle that is difficult to investigate in humans and challenging to treat. However, some progress is being made to enlighten the underlying maternal brain dysfunction, and recent evidence indicates that mothers' low responsiveness to their toddler correlates with less efficient oxytocin receptor gene variants [225]. Breaking the repeated cycle

of transgenerational poor parental care would have long lasting desirable emotional and economical benefits for individuals, families and governments. Simply targeting the dopamine/oxytocin systems is not a viable option. Since problems occur at multiple cellular, organismal and social levels, addressing each parameter on its own is not likely to impact on the problem in any meaningful way, and a multidimensional approach will be required.

A dysregulation of oxytocin and/or dopamine activity coupled with an impairment of social interactions has also been observed in a variety of diseases and disorders ranging from anorexia to Parkinson's disease [182]. Furthermore there has been an expansion in the research of an oxytocin or dopamine basis (and those neural circuitries upstream/downstream) in multiple behavioral syndromes. This review does not aim to cover the all diseases and disorders exhibiting inherent abnormalities in behavior but will now consider the role of these neuromodulators in selected behavioral disorders such as drug addiction and anorexia.

Addiction

The DSM-IV recently classified drug addiction as an individual who persists in the using of alcohol or other drugs despite problems related to use of the substance, resulting in a significant impairment in functioning. Due to the devastating impact on addicts' lives and the socio-economic burden associated with addiction, the neural basis of drug use disorders is being increasingly explored. Hurdles such as the heterogeneous nature of biological and genetic determinants add to the complexities of understanding drug seeking behavior. Additionally, drug addiction models used in experimental studies do not consider social environment and so do not closely mirror drug-seeking behavior in humans. Some elegant preclinical studies however, have revealed interesting information regarding social consequences of drug use [226,227] and those neural correlates, such as oxytocin, subserving addiction [228,229].

The role of the mesolimbic dopamine system in the rewarding effects of drugs of abuse has been well documented [230–233], with MDMA, cocaine and opiates known to influence dopaminergic neurotransmission in these motivational pathways [229,234] and produce marked impairment in prosocial behavior [235,236]. The mesolimbic system has been established as a key component of rewarding properties of natural rewards such as sex and food and maladaptive rewards such as drugs of abuse and is believed to drive chronic drug use. As mentioned previously, dopaminergic projections originating in the VTA and extending to the NA, amygdala,

olfactory tubercle and prefrontal cortex (see basic framework diagram in Figure 2, not all nuclei shown) comprise the mesolimbic pathway. In humans, fMRI studies revealed distinct brain activation in the VTA during cocaine self-administration in cocaine addicts which is indicative of mesolimbic dopamine activation [237]. Animal studies using *in vivo* microdialysis, where small changes in neurotransmitter concentrations can be detected, have shown that psychostimulants such as alcohol and cocaine activate mesolimbic reward circuits and increase dopamine levels in the NA [238,239]. Specific dopamine receptors mediating drug reinforcement pathways have yet to be elucidated; however, dopamine D1-like and D2-like receptors appear to be implicated in regulating the acute reinforcing properties of cocaine as evidenced from rodent studies where dopamine agonists and antagonists acting on all dopamine receptors reinstated or inhibited drug-seeking behavior, respectively [240,241]. Furthermore, disruption to the mesolimbic dopamine via selective ablation of the NA results in decreased self-administration of cocaine without affecting feeding behavior in rats [242]. Taken together, these findings emphasize the importance of the mesolimbic dopamine reward pathway in the processing of maladaptive rewards in particular, which drives continued drug administration leading to chronic drug abuse.

Because drug addiction has such a profound effect on a wide spectrum of social behaviors including social bonding, maternal and sexual behavior, it is not surprising that oxytocin has also become a key area of focus in addiction research. A role for oxytocin in drug addiction is not altogether surprising since oxytocin fibers innervate and receptors are expressed in known dopamine-containing nuclei important in reward assessment including the VTA and amygdala [79,80,82,243].

Dopamine/Oxytocin Interactions

Drugs of abuse known to target the mesolimbic system, such as cocaine markedly reduce the levels of oxytocin in the hippocampus, hypothalamus, NA and plasma when taken repeatedly over time [244]. These long-term disruptions to central oxytocin systems due to chronic abuse of psychostimulants, presumably contributes to the impaired social and emotional capabilities often observed in drug addicts. Whilst it is apparent that dopamine can influence oxytocin release in a chronic drug use context, these two neuromodulators may also function in a bidirectional manner during the development of tolerance and dependence to drugs of abuse.

It has been suggested that oxytocin may possess antipsychotic properties due to its ability to block cocaine-

induced dopamine release in the NA [245] and to attenuate characteristic locomotor activities associated with cocaine addiction [229,246–248]. Furthermore, microinjection of physiological doses of oxytocin into the NA, amygdala and the hippocampus attenuate morphine tolerance and dependence and cocaine-induced hyperactive locomotor activity [249]. So, the data suggest a potential role for oxytocin in the regulation of chronic drug abuse by influencing dopaminergic activity in key limbic brain sites and altering behavioral responses associated with addiction.

In addition to having a neuromodulatory role influencing neuroadaptive processes responsible for tolerance and dependence [229], recent findings have also alluded to a role for oxytocin in drug withdrawal [234,250]. Opiate drugs strongly inhibit oxytocin neurones since they coexpress opioid receptors. Endogenous opioid systems control oxytocin during physiological conditions, e.g., pregnancy and birth [27]. However, oxytocin neurones develop tolerance and dependence for opiates, which means that they are able to maintain their physiological roles even under pathologic conditions which may include drug addiction [251]. On the flip side, it also means that upon withdrawal and similar to dopamine systems [252,253], the whole oxytocin system is dysfunctional (temporarily or could be permanently) and this has long term physiological and social implications for those trying to withdraw and avoid drug taking situations or triggers.

The data suggest oxytocin action in key limbic brain regions has a regulatory role in attenuating drug tolerance and dependence and promoting drug withdrawal, presumably via its actions on mesolimbic dopamine reward pathways. So, referring to our original framework (Figure 2), there appears to be a growing body of evidence to suggest dopamine and oxytocin pathways may be two potential neural correlates mediating drug addiction. Central oxytocin sites are one area of addiction neurobiology that has yet to be fully explored and may serve as a potential neural substrate that could be potentially exploited for pharmacotherapeutic benefit in the treatment of drug use disorders and withdrawal.

Treatments for Addiction

Treatment for nicotine addiction has shown some success with the use of drugs such as bupropion (inhibits the depletion of central dopamine and noradrenaline stores and antagonizes nicotinic receptors) [254] and varenicline (mimics the actions of nicotine) which were both shown to promote smoking abstinence [255,256]. However, currently there are no available

pharmacotherapies, which specifically target other forms of psychostimulant addiction. Approved treatment options merely treat the symptoms associated with the addiction. Medication alone is not remotely sufficient to effectively tackle drug addiction. Psychotherapy, self-help/support groups and community-based projects such as substance abuse programs all play extremely important roles in trying to achieve and maintain abstinence in drug use disorders. In cocaine addiction there have been numerous clinical studies conducted and designed to target various neurotransmitter pathways in the brain with central GABA and dopamine systems proving to be potential therapeutic endpoints [257]. Potentiation of GABAergic neurotransmission via GABA_A and GABA_B receptor activation or inhibiting the degradation of GABA have reported some success in reducing cocaine use in dependent subjects [258–260]. In addition, drugs acting to enhance central dopaminergic activity (presumably in the mesolimbic system) have shown some success in humans as evidenced by an increase in urine screening protocols testing negative for addictive drugs [261]. The current medical therapy for opioid dependence is oral administration of the opioid analgesic, methadone. Methadone treatment remains the most effective treatment for opioid dependent patients, proving to be a more effective deterrent of heroin use than other nonpharmacological methods such as detoxification programs and prescribing placebo medication [262]. Whilst findings from controlled clinical trials using pharmacological agents to, for example, interfere with the metabolic degradation of alcohol or potentiate GABAergic function, have shown some promise in regard to amelioration of withdrawal symptoms and reducing drug seeking behavior [263,264]. The limited data available and pharmacologically induced adverse side effects emphasize the need for more exploratory work to be conducted in preclinical and clinical addiction studies. Preliminary work investigating the role of oxytocin in acute and long term repetitive drug use has implicated effectiveness of this neuropeptide in influencing neuro-adaptive processes and ultimately drug craving behavior [234]. Intranasal oxytocin administration would presumably help to redress hypothalamic oxytocin levels that are found to be diminished in alcohol dependent subjects [265]. Other therapies to restrain wildly high central oxytocin during opiate withdrawal are envisaged as potentially useful. Presumably oxytocin-induced stimulation of mesolimbic dopamine pathways to improve mood and behavioral symptoms associated with drug seeking behavior and withdrawal to stabilize social behaviors would be beneficial treatment strategies. Ultimately oxytocin's facilitatory role on psychosocial recovery may help to improve recovery and enhance receptivity to social support in dependent drug users.

Anorexia/Bulimia

The DSM-IV classification for eating disorders such as anorexia nervosa includes the refusal to maintain normal body weight, having an intense fear of gaining weight and a disturbance in the perception of body weight or shape. Anorexia nervosa and bulimia nervosa are complex psychological disorders associated with dysfunctional control of appetite for food and body weight. Anorexia involves fear-like and obsession-like behaviors, and affects about 0.3% of the population, mostly teenage girls. It is a DSM-IV classified mental disorder with serious comorbid consequences, including clinical depression, drug abuse and obsessive-compulsive disorder, as well as other related conditions like lack of reproductive ability. Bulimia is similar, though more common than anorexia, but also involves binge eating [266] and sufferers are particularly susceptible to addictive behaviors. Control of appetite is multidimensional, involving many interacting hypothalamic neuropeptides, peripheral energy signaling peptides and other neurotransmitters mediating hunger, satiety and reward, and evidently several of these elements are disturbed in anorexia and bulimia. The arcuate nucleus is a major integrating center for the control of appetite receiving input from higher centers and the gut as well as being able to sense circulating factors due to its leaky blood brain barrier. Major arcuate neuropeptides primarily include the orexigenic factors neuropeptide Y (NPY) and agouti-related protein (AgRP), and anorexigenic peptides such as alpha melanocyte stimulating hormone (α MSH), and cocaine- and amphetamine-related-transcript (CART). The arcuate communicates with the PVN and lateral hypothalamus in particular to control appetite, but also with other nuclei such as the SON, ventromedial nucleus and brainstem.

In fact the SON and PVN are emerging as regions that respond particularly to eating, neurones becoming activated strongly at the start of a meal [267]. Oxytocin is one of the nonarcuate anorexigenic peptides involved in signaling satiety, and so administration of oxytocin intracerebrally inhibits appetite, and oxytocin antagonist prolongs meal duration. Oxytocin is thought to be recruited downstream of the major appetite peptides from the arcuate nucleus [268]. For example, α MSH stimulates SON oxytocin neurones via melanocortin receptor 4 (MCR4). Interestingly, MCR4 agonists developed by the pharmaceutical industry to treat obesity also precipitated side effects such as prolonged penile erection in clinical trials, and we now know that is due to activation of central oxytocin systems [269], so there is a potential neuropeptide link between eating and sex. As well as the arcuate-oxytocin drive there is a bi-directional link

between the gut and oxytocin neurones, where eating (gastric distension) stimulates oxytocin activity in the SON and PVN [267] and oxytocin projections from the PVN to the brainstem regulate gastric reflexes [270]. However, apart from the brainstem, how and where oxytocin acts to mediate satiety is unclear.

A role for oxytocin in anorexia or bulimia is not well investigated. One might expect that if oxytocin dysregulation is involved in the low appetite element of anorexia that its expression and/or activity would be enhanced, but the evidence available tends to suggest that oxytocin is not an underlying cause [271]. Another anorexigenic peptide, nesfatin-1, is associated with anorexia. Intriguingly, nesfatin-1 is coexpressed in PVN oxytocin neurones [272] and stimulates intra-PVN oxytocin release. Since oxytocin mediates nesfatin-induced inhibition of appetite [273], this perhaps indicates a previously unrecognized role of oxytocin in anorexia, though further detailed studies are required. Evidence also seems to point towards mis-control of the orexigenic peptide AgRP in anorexia [274], and this might act at least partly via oxytocin too—AgRP acts as a MCR4 antagonist and inhibits oxytocin neurone responses to ingestive behavior [275]. On the other hand, lack of oxytocin may underlie decreased satiety in the Prader Willi syndrome [274], which is a rare genetic condition characterized by hyperphagia, obesity, reproduction failure and mental retardation. Clearly further research is needed to clarify a role for oxytocin in eating disorders.

Dopamine's function in signalling both food-seeking behavior and reward plays an important role in appetite and satiety [266,276]. First, dopamine deficient mice become completely aphagic and need dopamine replacement for any feeding to occur [277]. Secondly, mesocortical dopamine (from the VTA acting in the striatum and prefrontal cortex), but not mesolimbic dopamine (from VTA projecting to NA) pathways have a propensity to mediate food reward and anticipation of food [242,278]. Dopamine activity responds to olfactory and visual food-related stimuli and so in parallel with social recognition and sexual arousal responds to valid external cues. Dopamine is reported to be recruited by many appetite neuropeptides, including NPY, α MSH and AgRP, especially modulating NA dopamine release and activity [277]. The relevant dopaminergic regions, the VTA and NA, respond to gut signals such as ghrelin, which acts partly directly in the NA and VTA and partly indirectly via the arcuate, particularly AgRP [279] or α MSH, both of which also regulate oxytocin responses to appetite signals as indicated above. In addition, catecholamine lesion of the PVN disrupts AgRP expression and signalling, so upstream dopamine modulation of neuroendocrine systems

may also be recruited in control of the arcuate peptides [280].

Much evidence links dopamine with anorexia: there is altered dopamine function in the striatum and caudate putamen, along with other monoamines, particularly serotonin [266,281]. There is thought to be decreased dopamine turnover, as indicated from measuring CSF levels of dopamine metabolites in anorectic patients. This seems to reflect dysfunction of the dopaminergic system, which is associated with symptoms such as the inability to experience motivation and reward. Low dopamine is also associated with motor hyperactivity, a core symptom of anorexia nervosa that increases with the level of starvation [281]. PET imaging implicates altered dopamine activity in anorexic patients [266], while binge eating is associated with a polymorphism in the dopamine transporter gene in women [282], also indicating that altered dopamine concentrations at relevant target regions play an important role. Dysregulation of other monoamines is also apparent, with serotonin inhibiting food intake. However, dieting reduces tryptophan, the precursor for serotonin, perhaps explaining why patients have unexpectedly low serotonin in their CSF [281]. Serotonin and noradrenaline transmitter systems are both dysfunctional in bulimic patients [281], again based on CSF analysis so precision in changes at monoamine sources or targets are unknown.

With the lack of a relevant animal model for anorexia or bulimia, it is hard to find evidence in the literature for whether dopamine or oxytocin play a primary role in these eating disorders, let alone whether their interaction explains part of the phenotype. There have been limited opportunities to experimentally study any potential interaction between dopamine and oxytocin in this context, perhaps because the study of oxytocin in anorexia or obesity has been relatively limited up to now. However, if as indicated above there is a robust coregulation of reward by oxytocin and dopamine it might be expected that disturbances in food reward that are associated with anorexia/bulimia could be explained by disruption in oxytocin–dopamine pathways.

Treatment of Anorexia/Bulimia

In the past antidopaminergic drugs such as fluoxetine and phentermine proved effective for binge-eating disorders along with simultaneous cognitive behavioral therapy, but have not typically been used for treating anorexia despite the evidence for dopamine malfunction [281], probably due to poor tolerance of side effects. Anorexics seem to respond better to antidepressants and selective serotonin reuptake inhibitors than binge-eating patients [266]. SSRI are also effective—because anorexia is often

associated with depression-like symptoms, perhaps they effectively dissociate emotional elements such as anxiety from appetitive elements of the disorder. Given the role of central oxytocin in fear, anxiety and appetite control, as well as its known interactions with dopamine in other CNS disorders, oxytocin should be addressed more in the future as a potential component of treatment for these conditions. As indicated above, it is well recognized that anorexia/bulimia and other various behavioral disorders are associated with coexpression of anxiety- or depression-like symptoms so we will now address the roles of dopamine and oxytocin in major depression.

Major Depression

The DSM-IV definition of depression is the persistence of depressed mood and/or loss of interest or pleasure in daily activities consistently for at least a 2-week period. Depression is reported to affect up to 10% of people in the United Kingdom at some time during their lives, and one in fifty experience severe depression. Causes are multiple and varied and include acute and chronic conditions, as well as neonatal programming, which confers susceptibility to psychological disorders in later life, including depression. Furthermore, major depression can be experienced by mothers perinatally (postpartum psychosis), also with consequences for the child. In other disorders such as Parkinson's Disease, schizophrenia and anorexia, depression is comorbid due to the neurochemical nature of the condition. In addition, many CNS disorders elicit depression due to lack of support, and lack of recovery or appropriate treatment. Symptoms of chronic or major depression include feelings of sadness, low self-worth, withdrawal from social situations, high anxiety and poor stress coping.

A major cause of depression is chronic stress. A core hypothalamic center mediating stress responses is the hypothalamo-pituitary-adrenal (HPA) axis, where the expression, release and responsivity of corticotrophin-releasing hormone (CRH) and vasopressin from the PVN are altered, leading to prolonged misregulation of pituitary adrenocorticotrophic hormone (ACTH) and adrenal cortex glucocorticoids (cortisol in humans and corticosterone in rodents). Chronic stress leads to sustained characteristic changes in physiology and behavior that are typical of depression. The typical hypercortisolemia and elevated sympathetic hormone tone are due to stress-induced adaptations in the limbic and hypothalamic control centers, including altered glucocorticoid feedback control [283]. Although there are many brain and body adaptations with depression, and multiple interacting mechanisms have been proposed to either underlie or

arise as a consequence of depression, for brevity this review aims to focus only on potential mechanisms involving dopamine and oxytocin.

Stress, or other mechanisms arising from trauma or disease, causes dysregulation of central monoamines (serotonin, noradrenaline, and dopamine), which are major players mediating the adverse symptoms of depression [284]. Noradrenaline and serotonin act in a variety of relevant brain regions, including primarily in the PVN, but dopamine activity in the prefrontal cortex, NA, amygdala, and BNST, rather than the hypothalamus, is associated with stress response [285]. The main dopamine neurones that are stress sensitive are in the VTA and they project to cortical regions, so dopamine release increases in the prefrontal cortex in response to a variety of stressors [286]. Therefore stress effects on dopamine release and action in hypothalamic nuclei such as the PVN and SON are weak and dopamine's role in stress is mostly limited to hedonic and reward aspects of stress [286]. Analysis of human brain regions using microarray particularly reveals abnormal dopamine regulation in the prefrontal cortex in psychiatric disorders such as depression and bi-polar disorder [287]. Glutamate and GABA systems are also evidently dysregulated there, so there is a disintegration of the major excitatory and inhibitory neurotransmitters as well as the monoamines, and these modifications may be linked.

There are a variety of rodent models of depression and a suite of physiological and behavioral tests to analyse perceptible components of depressive symptoms. One such test is the dexamethasone-suppression test, which reveals alterations in glucocorticoid feedback. It has been shown using this approach that decreased dopamine in the prefrontal cortex accompanies prolonged stress-induced behavioral depression revealed [288], so the dopaminergic system is a potentially important target for therapies for human depression. Although it has been recognized for some time that low brain serotonin activity underlies HPA axis adaptations in depression, recent evidence further indicates that linked dysfunction of dopamine and serotonin systems is associated with depressive disorders [289–291].

As mentioned above (in the "Social Behavior" section), oxytocin has a strong antistress role, inhibiting HPA axis secretory and behavioral responses to stress, primarily by acting within the PVN to inhibit corticotrophin-releasing hormone neurones, although is also secreted and may inhibit pituitary corticotrophs [166]. Oxytocin also plays an important role in mediating anxiety and oxytocin-null mice exhibit increased anxiety behavior [166] by acting in the PVN, SON, and amygdala. Glucocorticoids such as corticosterone inhibit magnocellular oxytocin neurones [292], so during chronic stress and depression the

anxiolytic and antistress effects of oxytocin are attenuated, preventing action of this important controller, and so lack of central oxytocin is associated with depression.

In humans there is some evidence that low oxytocin correlates with depression in women, since plasma concentrations are reduced, but this was not observed in men [293]. If reduced oxytocin secretion reflects reduced central oxytocin release in women it may indicate that the oxytocin system is susceptible to stress in women particularly and therefore could have selective consequences for reproduction and offspring care and development. In rat models of high anxiety oxytocin mediates enhanced maternal behavioral and hormonal responses to stress [294], perhaps in an effort to overcome the inherent anxiety phenotype. Because glucocorticoid secretion increases prior to birth in both rodents and women, and oxytocin neurone responses to stress are inhibited in late gestation [295], oxytocin dysfunction in the maternal rat brain is a candidate for explaining postnatal depression [294]. Oxytocin neurones are also sensitive to the rapidly changing sex steroid environment perinatally, further reinforcing the concept that oxytocin neurones participate in the onset of depressive symptoms. Furthermore, central oxytocin control of stress responsiveness may also be partly inherited since the important brain oxytocin receptor expression levels and patterns are susceptible to epigenetic inheritance [211,296], (see above).

Despite the multiple studies showing separate roles for dopamine and oxytocin, reports of a joint role for dopamine and oxytocin in depression are limited and tend to be negative, e.g., the dopamine agonist apomorphine has no effect on peripheral oxytocin secretion in depressed patients [297]. This may be since the main regions where dopamine responds to stress (i.e., prefrontal cortex) overlap little with oxytocin sources or targets, although oxytocin can act in the VTA, possibly on dopamine somata. With chronic stress or hypercortisolemia any drive to oxytocin neurones would be limited, preventing any protective effect of oxytocin or dopamine against depression. Therefore, referring to our framework, a lack of dopamine–oxytocin interaction in depression under typical conditions is suspected. On the other hand, it might be speculated that in physiological states where the oxytocin system is highly activated, such as at birth, that increased oxytocin action in the PVN, SON, amygdala, and BNST would impact more on dopamine signalling and/or rise above a threshold that activates VTA dopamine neurones. Unlike dopamine, strong evidence points towards an anxiolytic effect of oxytocin via serotonin in a mouse model of depression [291]. Thus oxytocin could still be a relevant target for therapeutic intervention in anxiety and depression disorders.

Treatments for Depression

Current drugs for treating depression include the tricyclic antidepressants, the selective serotonin reuptake inhibitors, monoamine oxidase inhibitors, and noradrenaline reuptake inhibitors. However more recent developments have shown the potential for dopamine reuptake inhibitor drugs, either alone or in combination with serotonin and/or noradrenaline uptake inhibitors, in treating depression and major depressive disorders [298,299]. Rat models and current clinical trials (clinicaltrials.gov) indicate that dopamine agonists are also effective antidepressants, acting partly via serotonin mechanisms [300]. However, oxytocin is not a pharmacological target as yet, although intranasal oxytocin treatment has the potential for reducing depression-related symptoms in men [140]. Since the current main drug treatment for depression, selective serotonin reuptake inhibitors, is not effective in a large majority of patients or has only a delayed effect in ameliorating symptoms [301], perhaps the interaction between oxytocin–serotonin, or oxytocin–dopamine–serotonin could be exploited further in developing new treatments for major depression.

Conclusion

Dopamine and oxytocin are two key centrally acting agents with widespread functions in the brain. Many behavioral disorders discussed are associated with oxytocin and/or dopamine dysregulation (Table 3). Whilst they are both involved in mediating organic functions such as penile erection, disruptions to either one of these pathways can have a marked effect on downstream neural processes, which can lead to profound social behavior deficits and establishment of altered behavioral states (e.g., social withdrawal and chronic drug use). We find dopamine and oxytocin may operate in a bidirectional manner driving organic functions such as penile erection (dopamine–oxytocin) and reinforcing/rewarding properties of social and addictive behaviors (oxytocin–dopamine) (see Figure 4). These two neuromodulators serve as potential neural correlates, which form a much larger neural network comprised of multiple neurochemical pathways and intricate circuitries. Trying to delineate a link between dopamine and oxytocin in normal and pathologic contexts remains a huge undertaking; however, progress is being made and warrants further and more thorough investigation.

Conflict of Interest

There were no conflicts of interest in preparation of this manuscript.

Table 3 Overview of the involvement of oxytocin and dopamine in CNS disorders and whether they are drug targets currently used in clinical therapy

Disorders	Oxytocin role	Dopamine role	Treatment targets? (dopamine/oxytocin?)
Sexual dysfunction			
erectile dysfunction- male	Decrease	Decrease	Dopamine or oxytocin
sexual desire disorders- female	Unknown	Unknown	Neither
Social dysfunction			
parental behavior	Decrease?	Unknown	Neither
autism	Decrease?	Decrease?	Dopamine
Drug addiction	Decrease	Increases ^a	Dopamine ^b or oxytocin ^b
Depression	Decrease	Decrease	Dopamine ^b
Eating disorders			
anorexia and bulimia	No change?	Increase	Dopamine ^c

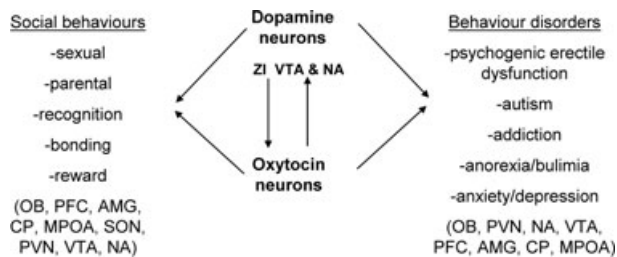
^aAnd/or mimic effects.^bPreliminary work/trials.^cIn the past but superseded by newer treatments.

Figure 4 Summary of dopamine and oxytocin involvement in social behavior and behavioral disorders. Summary of potential dopamine and oxytocin interactions underlying socio-affiliative behaviors and subsequent behavioral disorders. Central oxytocin neurons activated by in-certhypothalamic (ZI) dopamine input and mesolimbic dopamine pathways driven by hypothalamic and limbic oxytocin release comprise part of the neural circuitry governing social behaviors. Disruptions or changes in these neurochemical pathways may partly underpin pathophysiologic mechanisms contributing to organic functions such as erectile dysfunction, but also adversely affect an array of social parameters, which can lead to the development of profound behavioral disorders. NA, nucleus accumbens; ZI, MPOA, medial preoptic nucleus; PVN, paraventricular nucleus; SON, supraoptic nucleus; AMG, amygdala; VTA, ventral tegmental area; HC, hippocampus; OB, olfactory bulbs; CP, caudate putamen; PFC, prefrontal cortex.

Acknowledgments

T.A.B. was funded by a BBSRC CASE award and Pfizer Pharmaceutical Ltd as a postgraduate student. A.J.D. is supported by The Wellcome Trust and the MRC.

References

1. Fineberg NA, Krishnaiah RB, Moberg J, O'Doherty C. Clinical screening for obsessive-compulsive and related disorders. *Israel J Psychiatry Relat Sci* 2008;**45**:151–163.

2. Fink M, Akimova E, Spindelegger C, Hahn A, Lanzenberger R, Kasper S. Social anxiety disorder: Epidemiology, biology and treatment. *Psychiatr Danubina* 2009;**21**:533–542.
3. Cousins DA, Butts K, Young AH. The role of dopamine in bipolar disorder. *Bipolar Disord* 2009;**11**:787–806.
4. Gimpl G, Fahrenholz F. The oxytocin receptor system: Structure, function, and regulation. *Physiol Rev* 2001;**81**:629–683.
5. Lee HJ, Macbeth AH, Pagani JH, Young WS. Oxytocin: The great facilitator of life. *Prog Neurobiol* 2009;**88**:127–151.
6. Bora E, Yucel M, Allen NB. Neurobiology of human affiliative behaviour: Implications for psychiatric disorders. *Curr Opin Psychiatry* 2009;**22**:320–325.
7. Den Oudsten BL, Van Heck GL, De Vries J. Quality of life and related concepts in Parkinson's disease: A systematic review. *Movement Disord* 2007;**22**:1528–1537.
8. Roberts DL, Penn DL. Social cognition and interaction training (SCIT) for outpatients with schizophrenia: A preliminary study. *Psychiatry Res* 2009;**166**:141–147.
9. Argiolas A, Melis MR. The role of oxytocin and the paraventricular nucleus in the sexual behaviour of male mammals. *Physiol Behav* 2004;**83**:309–317.
10. Argiolas A, Melis MR. Central control of penile erection: Role of the paraventricular nucleus of the hypothalamus. *Prog Neurobiol* 2005;**76**:1–21.
11. Skuse DH, Gallagher L. Dopaminergic-neuropeptide interactions in the social brain. *Trends Cogn Sci* 2009;**13**:27–35.
12. Sanghera MK, Anselmofranci J, Mccann SM. Effect of medial zona incerta lesions on the ovulatory surge of gonadotropins and prolactin in the rat. *Neuroendocrinology* 1991;**54**:433–438.
13. Bitran D, Hull EM, Holmes GM, Lookingland KJ. Regulation of Male-rat copulatory-behavior by preoptic

- incertoypothalamic dopamine neurons. *Brain Res Bull* 1988;**20**:323–331.
14. Eriksson J, Olausson B, Jankowska E. Antispastic effects of L-dopa. *Exp Brain Res* 1996;**111**:296–304.
 15. Skoog B, Noga BR. Dopaminergic control of transmission from group-Ii muscle afferents to spinal neurons in the cat and Guinea-Pig. *Exp Brain Res* 1995;**105**:39–47.
 16. Holstege JC, VanDijken H, Buijs RM, Goedknecht H, Gosens T, Bongers CMH. Distribution of dopamine immunoreactivity in the rat, cat, and monkey spinal cord. *J Comparat Neurol* 1996;**376**:631–652.
 17. Ariano MA, Monsma FJ, Barton AC, Kang HC, Haugland RP, Sibley DR. Direct visualization and cellular-localization of D1 and D2 dopamine-receptors in rat forebrain by use of fluorescent ligands. *Proc Natl Acad Sci USA* 1989;**86**:8570–8574.
 18. Defagot MC, Malchiodi EL, Villar MJ, Antonelli MC. Distribution of D4 dopamine receptor in rat brain with sequence-specific antibodies. *Mol Brain Res* 1997;**45**:1–12.
 19. Khan ZU, Gutierrez A, Martin R, Penafiel A, Rivera A, De La Calle A. Differential regional and cellular distribution of dopamine D2-like receptors: An immunocytochemical study of subtype-specific antibodies in rat and human brain. *J Comparat Neurol* 1998;**402**:353–371.
 20. Khan ZU, Gutierrez A, Martin R, Penafiel A, Rivera A, De La Calle A. Dopamine D5 receptors of rat and human brain. *Neuroscience* 2000;**100**:689–699.
 21. Mansour A, Meadorwoodruff JH, Bunzow JR, Civelli O, Akil H, Watson SJ. Localization of dopamine-D2 receptor messenger-Rna and D1 and D2 receptor-binding in the rat-brain and pituitary—An insitu hybridization-receptor autoradiographic analysis. *J Neurosci* 1990;**10**:2587–2600.
 22. Levey AI, Hersch SM, Rye DB, et al. Localization of D(1) and D(2) dopamine-receptors in brain with subtype-specific antibodies. *Proc Natl Acad Sci USA* 1993;**90**:8861–8865.
 23. Campbell A. Attachment, aggression and affiliation: The role of oxytocin in female social behavior. *Biol Psychol* 2008;**77**:1–10.
 24. Cushing BS, Carter CS. Peripheral pulses of oxytocin increase partner preferences in female, but not male, prairie voles. *Horm Behav* 2000;**37**:49–56.
 25. Ferguson JN, Young LJ, Hearn EF, Matzuk MM, Insel TR, Winslow JT. Social amnesia in mice lacking the oxytocin gene. *Nat Genet* 2000;**25**:284–288.
 26. Kendrick KM. Oxytocin, motherhood and bonding. *Exp Physiol* 2000;**85**:111S–124S.
 27. Russell JA, Leng G, Douglas AJ. The magnocellular oxytocin system, the fount of maternity: Adaptations in pregnancy. *Front Neuroendocrinol* 2003;**24**:27–61.
 28. Douglas AJ, Johnstone LE, Leng G. Neuroendocrine mechanisms of change in food intake during pregnancy: A potential role for brain oxytocin. *Physiol Behav* 2007;**91**:352–365.
 29. Bale TL, Davis AM, Auger AP, Dorsa DM, McCarthy MM. CNS region-specific oxytocin receptor expression: Importance in regulation of anxiety and sex behavior. *J Neurosci* 2001;**21**:2546–2552.
 30. McCarthy MM, McDonald CH, Brooks PJ, Goldman D. An anxiolytic action of oxytocin is enhanced by estrogen in the mouse. *Physiol Behav* 1996;**60**:1209–1215.
 31. Ring RH, Malberg JE, Potestio L, et al. Anxiolytic-like activity of oxytocin in male mice: Behavioral and autonomic evidence, therapeutic implications. *Psychopharmacology* 2006;**185**:218–225.
 32. Insel TR, Young L, Wang ZX. Central oxytocin and reproductive behaviours. *Rev Reprod* 1997;**2**:28–37.
 33. Russell JA, Douglas AJ, Ingram CD. Brain preparations for maternity-adaptive changes in behavioral and neuroendocrine systems during pregnancy and lactation. An overview. *Prog Brain Res* 2001;**133**:1–38.
 34. Neumann ID. Alterations in behavioral and neuroendocrine stress coping strategies in pregnant, parturient and lactating rats. *Prog Brain Res* 2001;**133**:143–152.
 35. Tang Y, Rampin O, Calas A, Facchinetti P, Giuliano F. Oxytocinergic and serotonergic innervation of identified lumbosacral nuclei controlling penile erection in the male rat. *Neuroscience* 1998;**82**:241–254.
 36. Badoer E. Hypothalamic paraventricular nucleus and cardiovascular regulation. *Clin Exp Pharmacol Physiol* 2001;**28**:95–99.
 37. Mack SO, Kc P, Wu M, Coleman BR, Tolentino-Silva FP, Haxhiu MA. Paraventricular oxytocin neurons are involved in neural modulation of breathing. *J Appl Physiol* 2002;**92**:826–834.
 38. Petersson M. Cardiovascular effects of oxytocin. *Prog Brain Res* 2002;**139**:281–288.
 39. Sawchenko PE, Swanson LW. Relationship of oxytocin pathways to the control of neuroendocrine and autonomic function. *J Steroid Biochem Mol Biol* 1984;**20**:87–103.
 40. Sofroniew MV, Weindl A. Extrahypothalamic neurophysin-containing perikarya, fiber pathways and fiber clusters in rat-brain. *Endocrinology* 1978;**102**:334–337.
 41. Argiolas A. Neuropeptides and sexual behaviour. *Neurosci Biobehav Rev* 1999;**23**:1127–1142.
 42. Insel TR. A neurobiological basis of social attachment. *Am J Psychiatry* 1997;**154**:726–735.
 43. Kendrick KM. The neurobiology of social bonds. *J Neuroendocrinol* 2004;**16**:1007–1008.
 44. Richard P, Moos F, Freundmercier MJ. Central effects of oxytocin. *Physiol Rev* 1991;**71**:331–370.
 45. Moos F, Freundmercier MJ, Guerne Y, Guerne JM, Stoeckel ME, Richard P. Release of oxytocin and

- vasopressin by magnocellular nuclei invitro—Specific facilitatory effect of oxytocin on its own release. *J Endocrinol* 1984;**102**:63–72.
46. Moos F, Poulain DA, Rodriguez F, Guerne Y, Vincent JD, Richard P. Release of oxytocin within the supraoptic nucleus during the milk ejection reflex in rats. *Exp Brain Res* 1989;**76**:593–602.
 47. Neumann I, Ludwig M, Engelmann M, Pittman QJ, Landgraf R. Simultaneous microdialysis in blood and brain—Oxytocin and vasopressin release in response to central and peripheral osmotic stimulation and suckling in the rat. *Neuroendocrinology* 1993;**58**:637–645.
 48. Neumann I, Russell JA, Landgraf R. Oxytocin and vasopressin release within the supraoptic and paraventricular nuclei of pregnant, parturient and lactating rats—A microdialysis study. *Neuroscience* 1993;**53**:65–75.
 49. Ludwig M, Sabatier N, Dayanithi G, Russell JA, Leng G. The active role of dendrites in the regulation of magnocellular neurosecretory cell behavior. *Vasopressin Oxytocin: Genes Clinl Appl* 2002;**139**:247–256.
 50. Ludwig M, Leng G. Dendritic peptide release and peptide-dependent behaviours. *Nat Rev Neurosci* 2006;**7**:126–136.
 51. Neumann I, Russell JA, Landgraf R. Oxytocin and vasopressin release within the supraoptic and paraventricular nuclei of pregnant, parturient and lactating rats—A microdialysis study. *Neuroscience* 1993;**53**:65–75.
 52. Bosch OJ, Kromer SA, Brunton PJ, Neumann ID. Release of oxytocin in the hypothalamic paraventricular nucleus, but not central amygdala or lateral septum in lactating residents and virgin intruders during maternal defence. *Neuroscience* 2004;**124**:439–448.
 53. Neumann ID, Toschi N, Ohl F, Torner L, Kromer SA. Maternal defence as an emotional stressor in female rats: Correlation of neuroendocrine and behavioural parameters and involvement of brain oxytocin. *Eur J Neurosci* 2001;**13**:1016–1024.
 54. Engelmann M, Ebner K, Landgraf R, Holsboer F, Wotjak CT. Emotional stress triggers intrahypothalamic but not peripheral release of oxytocin in male rats. *J Neuroendocrinol* 1999;**11**:867–872.
 55. Zhang XH, Filippi S, Vignozzi L, et al. Identification, localization and functional in vitro and in vivo activity of oxytocin receptor in the rat penis. *J Endocrinol* 2005;**184**:567–576.
 56. Freundmercier MJ, Stoeckel ME, Klein MJ. Oxytocin receptors on oxytocin neurons—Histoautoradiographic detection in the lactating rat. *J Physiol* 1994;**480**:155–161.
 57. Yoshimura R, Kiyama H, Kimura T, Araki T, Maeno H, Tanizawa O, Tohyama M. Localization of oxytocin receptor messenger-ribonucleic-acid in the rat-brain. *Endocrinology* 1993;**133**:1239–1246.
 58. Veronneau-Longueville F, Rampin O, Freund-Mercier MJ, et al. Oxytocinergic innervation of autonomic nuclei controlling penile erection in the rat. *Neuroscience* 1999;**93**:1437–1447.
 59. Ferguson JN, Aldag JM, Insel TR, Young LJ. Oxytocin in the medial amygdala is essential for social recognition in the mouse. *J Neurosci* 2001;**21**:8278–8285.
 60. Huber D, Veinante P, Stoop R. Vasopressin and oxytocin excite distinct neuronal populations in the central amygdala. *Science* 2005;**308**:245–248.
 61. Terenzi MG, Ingram CD. Oxytocin-induced excitation of neurones in the rat central and medial amygdaloid nuclei. *Neuroscience* 2005;**134**:345–354.
 62. Baskerville TA, Douglas AJ. Interactions between dopamine and oxytocin in the control of sexual behaviour. *Prog Brain Res* 2008;**170**:277–290.
 63. Wang ZX, Aragona BJ. Neurochemical regulation of pair bonding in male prairie voles. *Physiol Behav* 2004;**83**:319–328.
 64. Buijs RM, Geffard M, Pool CW, Hoorneman EMD. The dopaminergic innervation of the supraoptic and paraventricular nucleus—A light and electron microscopical study. *Brain Res* 1984;**323**:65–72.
 65. Decavel C, Geffard M, Calas A. Comparative-study of dopamine-immunoreactive and noradrenaline-immunoreactive terminals in the paraventricular and supraoptic nuclei of the rat. *Neurosci Lett* 1987;**77**:149–154.
 66. Smeltzer MD, Curtis JT, Aragona BJ, Wang ZX. Dopamine, oxytocin, and vasopressin receptor binding in the medial prefrontal cortex of monogamous and promiscuous voles. *Neurosci Lett* 2006;**394**:146–151.
 67. Pyner S. Neurochemistry of the paraventricular nucleus of the hypothalamus: Implications for cardiovascular regulation. *J Chem Neuroanat* 2009;**38**:197–208.
 68. Ross HE, Cole CD, Smith Y, Neumann ID, Landgraf R, Murphy AZ, Young LJ. Characterization of the oxytocin system regulating affiliative behavior in female prairie voles. *Neuroscience* 2009;**162**:892–903.
 69. Schwartz JC, Diaz J, Griffon N, Levesque D, Martres MP, Sokoloff P. Multiple dopamine receptors: The D3 receptor and actions of substances of abuse. *EXS* 1994;**71**:81–92.
 70. Baskerville TA, Allard J, Wayman C, Douglas AJ. Dopamine-oxytocin interactions in penile erection. *Eur J Neurosci* 2009;**30**:2151–2164.
 71. Melis MR, Succu S, Mascia MS, Cortis L, Argiolas A. Extra-cellular dopamine increases in the paraventricular nucleus of male rats during sexual activity. *Eur J Neurosci* 2003;**17**:1266–1272.
 72. Waldherr M, Neumann ID. Centrally released oxytocin mediates mating-induced anxiolysis in male rats. *Proc Natl Acad Sci USA* 2007;**104**:16681–16684.
 73. Melis MR, Melis T, Cocco C, et al. Oxytocin injected into the ventral tegmental area induces penile erection and

- increases extracellular dopamine in the nucleus accumbens and paraventricular nucleus of the hypothalamus of male rats. *Eur J Neurosci* 2007;**26**:1026–1035.
74. Melis MR, Succu S, Sanna F, Boi A, Argiolas A. Oxytocin injected into the ventral subiculum or the posteromedial cortical nucleus of the amygdala induces penile erection and increases extracellular dopamine levels in the nucleus accumbens of male rats. *Eur J Neurosci* 2009;**30**:1349–1357.
 75. Giuliano F, Allard J. Dopamine and sexual function. *Int J Impotence Res* 2001;**13**:S18–S28.
 76. Young KA, Liu Y, Wang ZX. The neurobiology of social attachment: A comparative approach to behavioral, neuroanatomical, and neurochemical studies. *Comp Biochem Physiol C-Toxicol Pharmacol* 2008;**148**:401–410.
 77. Paredes RG, Agmo A. Has dopamine a physiological role in the control of sexual behavior? A critical review of the evidence. *Prog Neurobiol* 2004;**73**:179–226.
 78. Pattij T, de Jong TR, Uitterdijk A, et al. Individual differences in male rat ejaculatory behaviour: Searching for models to study ejaculation disorders. *Eur J Neurosci* 2005;**22**:724–734.
 79. Buijs RM. Intra-hypothalamic and extrahypothalamic vasopressin and oxytocin pathways in rat—Pathways to limbic system, medulla-oblongata and spinal-cord. *Cell Tissue Res* 1978;**192**:423–435.
 80. Sofroniew MV. Morphology of vasopressin and oxytocin neurons and their central and vascular projections. *Prog Brain Res* 1983;**60**:101–114.
 81. Roeling TAP, Veening JG, Peters JPW, Vermelis MEJ, Nieuwenhuys R. Efferent connections of the hypothalamic grooming area in the rat. *Neuroscience* 1993;**56**:199–225.
 82. Freundmercier MJ, Stoeckel ME, Palacios JM, Pazos A, Reichhart JM, Porte A, Richard P. Pharmacological characteristics and anatomical distribution of [H-3] oxytocin-binding sites in the Wistar rat-brain studied by autoradiography. *Neuroscience* 1987;**20**:599–614.
 83. Vaccari C, Lolait SJ, Ostrowski NL. Comparative distribution of vasopressin V1b and oxytocin receptor messenger ribonucleic acids in brain. *Endocrinology* 1998;**139**:5015–5033.
 84. Swanson LW. The projections of the ventral tegmental area and adjacent regions—A combined fluorescent retrograde tracer and immunofluorescence study in the rat. *Brain Res Bull* 1982;**9**:321–353.
 85. Fallon JH, Loughlin SE. Substantia nigra. In: Paxinos G, editor. *The rat nervous system*. New York: Academic Press, 1995.
 86. Aragona BJ, Wang ZX. The prairie vole (*Microtus ochrogaster*): An animal model for behavioral neuroendocrine research on pair bonding. *Ilar J* 2004;**45**:35–45.
 87. Gingrich JA, Dearry A, Falardeau P, Bates MD, Freneau RT, Caron MG. Location and molecular-cloning of D1 dopamine receptor. *Neurochem Int* 1992;**20**:S9–S15.
 88. Young LJ. Oxytocin and vasopressin receptors and species-typical social behaviors. *Horm Behav* 1999;**36**:212–221.
 89. Liu Y, Wang ZX. Nucleus accumbens oxytocin and dopamine interact to regulate pair bond formation in female prairie voles. *Neuroscience* 2003;**121**:537–544.
 90. Safarinejad MR, Hosseini S. Erectile dysfunction: Clinical guidelines (1). *Urol J* 2004;**1**:133–147.
 91. Zhang XH, Hu LQ, Yin J, Mo ZN, Chen J. Rat model of erectile dysfunction caused by cavernous nerve ablation. *Chin Med J* 2002;**115**:1179–1182.
 92. Clement P, Peeters M, Bernabe J, Denys P, Alexandre L, Giuliano F. Brain oxytocin receptors mediate ejaculation elicited by 7-hydroxy-2-(di-N-propylamino) tetralin (7-OH-DPAT) in anaesthetized rats. *Br J Pharmacol* 2008;**154**:1150–1159.
 93. de Jong TR, Veening JG, Olivier B, Waldinger MD. Oxytocin involvement in SSRI-induced delayed ejaculation: A review of animal studies. *J Sexual Med* 2007;**4**:14–28.
 94. Andersson KE. Neurophysiology/pharmacology of erection. *Int J Impotence Res* 2001;**13**:S8–S17.
 95. Giuliano F, Rampin O. Central neural regulation of penile erection. *Neurosci Biobehav Rev* 2000;**24**:517–533.
 96. Heaton JPW. Central neuropharmacological agents and mechanisms in erectile dysfunction: The role of dopamine. *Neurosci Biobehav Rev* 2000;**24**:561–569.
 97. Ralph DJ. Normal erectile function. *Clin Cornerstone* 2005;**7**:13–18.
 98. Carmichael MS, Humbert R, Dixen J, Palmisano G, Greenleaf W, Davidson JM. Plasma oxytocin increases in the human sexual-response. *J Clin Endocrinol Metabol* 1987;**64**:27–31.
 99. Uckert S, Becker AJ, Ness BO, Stief CG, Scheller F, Knapp WH, Jonas U. Oxytocin plasma levels in the systemic and cavernous blood of healthy males during different penile conditions. *World J Urol* 2003;**20**:323–326.
 100. Arletti R, Bazzani C, Castelli M, Bertolini A. Oxytocin improves male copulatory performance in rats. *Horm Behav* 1985;**19**:14–20.
 101. Martino B, Hsieh GC, Hollingsworth PR, Mikusa JP, Moreland RB, Bitner RS. Central oxytocinergic and dopaminergic mechanisms regulating penile erection in conscious rats. *Pharmacol Biochem Behav* 2005;**81**:797–804.
 102. Miner MM, Seftel AD. Centrally acting mechanisms for the treatment of male sexual dysfunction. *Urol Clin North Am* 2007;**34**:483–496, v.
 103. Hull EM, Dominguez JM. Sexual behavior in male rodents. *Horm Behav* 2007;**52**:45–55.
 104. Melis MR, Succu S, Mascia MS, Argiolas A. PD-168077, a selective dopamine D-4 receptor agonist, induces

- penile erection when injected into the paraventricular nucleus of male rats. *Neurosci Lett* 2005;**379**:59–62.
105. Melis MR, Succu S, Sanna F, et al. PIP3EA and PD-168077, two selective dopamine D4 receptor agonists, induce penile erection in male rats: Site and mechanism of action in the brain. *Eur J Neurosci* 2006;**24**:2021–2030.
 106. Caquineau C, Leng G, Guan XMM, Jiang M, Van Der Ploeg L, Douglas AJ. Effects of alpha-melanocyte-stimulating hormone on magnocellular oxytocin neurones and their activation at intromission in male rats. *J Neuroendocrinol* 2006;**18**:685–691.
 107. Bridges TE, Hillhouse EW, Jones MT. Effect of dopamine on neurohypophyseal hormone-release *in vivo* and from rat neural lobe and hypothalamus *in vitro*. *J Physiol* 1976;**260**:647–666.
 108. Melis MR, Stancampiano R, Argiolas A. Hippocampal oxytocin mediates apomorphine-induced penile erection and yawning. *Pharmacol Biochem Behav* 1992;**42**:61–66.
 109. Cameron JL, Pomerantz SM, Layden LM, Amico JA. Dopaminergic stimulation of oxytocin concentrations in the plasma of male and female monkeys by apomorphine and a D2 receptor agonist. *J Clin Endocrinol Metabol* 1992;**75**:855–860.
 110. Succu S, Sanna F, Melis T, Boi A, Argiolas A, Melis MR. Stimulation of dopamine receptors in the paraventricular nucleus of the hypothalamus of male rats induces penile erection and increases extra-cellular dopamine in the nucleus accumbens: Involvement of central oxytocin. *Neuropharmacology* 2007;**52**:1034–1043.
 111. Melis MR, Argiolas A, Stancampiano R, Gessa GL. Effect of apomorphine on oxytocin concentrations in different brain-areas and plasma of male-rats. *Eur J Pharmacol* 1990;**182**:101–107.
 112. Mason WT. Excitation by dopamine of putative oxytocinergic neurones in the rat supraoptic nucleus *in vitro*: Evidence for two classes of continuously firing neurones. *Brain Res* 1983;**267**:113–121.
 113. Yang CR, Bourque CW, Renaud LP. Dopamine-D2 receptor activation depolarizes rat supraoptic neurons in hypothalamic explants. *J Physiol* 1991;**443**:405–419.
 114. Argiolas A, Melis MR, Mauri A, Gessa GL. Paraventricular nucleus lesion prevents yawning and penile erection induced by apomorphine and oxytocin but not by acth in rats. *Brain Res* 1987;**421**:349–352.
 115. Hawthorn J, Ang VTY, Jenkins JS. Effects of lesions in the hypothalamic paraventricular, supraoptic and suprachiasmatic nuclei on vasopressin and oxytocin in rat-brain and spinal-cord. *Brain Res* 1985;**346**:51–57.
 116. Melis MR, Stancampiano R, Argiolas A. Effect of excitatory amino-acid receptor antagonists on apomorphine-induced, oxytocin-induced and acth-induced penile erection and yawning in male-rats. *Eur J Pharmacol* 1992;**220**:43–48.
 117. Melis MR, Succu S, Iannucci U, Argiolas A. Oxytocin increases nitric oxide production in the paraventricular nucleus of the hypothalamus of male rats: Correlation with penile erection and yawning. *Regul Peptides* 1997;**69**:105–111.
 118. Witt DM, Insel TR. Increased Fos expression in oxytocin neurons following masculine sexual-behavior. *J Neuroendocrinol* 1994;**6**:13–18.
 119. Kita I, Yoshida Y, Nishino S. An activation of parvocellular oxytocinergic neurons in the paraventricular nucleus in oxytocin-induced yawning and penile erection. *Neurosci Res* 2006;**54**:269–275.
 120. Sermasi E, Coote JH. Oxytocin acts at V-1 receptors to excite sympathetic preganglionic neurons in neonate rat spinal-cord *in-vitro*. *Brain Res* 1994;**647**:323–332.
 121. Yang Z, Han DD, Coote JH. Cardiac sympatho-excitatory action of PVN-spinal oxytocin neurones. *Autonom Neurosci-Basic Clin* 2009;**147**:80–85.
 122. Succu S, Sanna F, Cocco C, et al. Oxytocin induces penile erection when injected into the ventral tegmental area of male rats: Role of nitric oxide and cyclic GMP. *Eur J Neurosci* 2008;**28**:813–821.
 123. Montague DK. Nonpharmacologic treatment of erectile dysfunction. *Rev Urol* 2002;**4**:9–16.
 124. Toda N, Ayajiki K, Okamura T. Nitric oxide and penile erectile function. *Pharmacol Therap* 2005;**106**:233–266.
 125. Altwein JE, Keuler FU. Oral treatment of erectile dysfunction with apomorphine SL. *Urol Int* 2001;**67**:257–263.
 126. Dinsmore W. Treatment of erectile dysfunction. *Int J STD AIDS* 2004;**15**:215–221.
 127. Tan DQ, Yao Y, Zhang J. Apomorphine and erectile dysfunction. *Natl J Androl* 2007;**13**:818–821.
 128. Behl CR, Pimplaskar HK, Sileno AP, deMeireles J, Romeo VD. Effects of physicochemical properties and other factors on systemic nasal drug delivery. *Adv Drug Deliv Rev* 1998;**29**:89–116.
 129. Lu W, Jiang WM, Chen J, Yin M, Wang ZJ, Jiang XG. Modulation of brain delivery and copulation by intranasal apomorphine hydrochloride. *Int J Pharmaceut* 2008;**349**:196–205.
 130. Kendirci M, Hellstrom WJG. Intranasal apomorphine Nastech pharmaceutical. *Idrugs* 2004;**7**:483–488.
 131. Smith JG, Merrill DC. Oxytocin for induction of labor. *Clin Obstetr Gynecol* 2006;**49**:594–608.
 132. Hellstrom WJG. Clinical applications of centrally acting agents in male sexual dysfunction. *Int J Impotence Res* 2008;**20**:S17–S23.
 133. Safarinejad MR, Hosseini SY. Salvage of sildenafil failures with bremelanotide: A randomized, double-blind, placebo controlled study. *J Urol* 2008;**179**:1066–1071.
 134. Sperling H, Lummen G, Schneider T, Rubben H. New treatment options for erectile dysfunction.

- Pharmacologic and nonpharmacologic options. *Herz* 2003;**28**:314–324.
135. Afif-Abdo J, Teloken C, Damiao R, et al. Comparative cross-over study of sildenafil and apomorphine for treating erectile dysfunction. *BJU Int* 2008;**102**:829–834.
 136. Lundberg PO, Ertekin C, Ghezzi A, et al. Neurosexology—Guidelines for neurologists. *Eur J Neurol* 2001;**8**:2–24.
 137. Wyllie MG. Erectile response with vardenafil in sildenafil nonresponders: A multicentre, double-blind, 12-week, flexible-dose, placebo-controlled erectile dysfunction clinical trial—Comment. *BJU Int* 2004;**94**:1309.
 138. Chini B, Manning M. Agonist selectivity in the oxytocin/vasopressin receptor family: New insights and challenges. *Biochem Soc Trans* 2007;**35**:737–741.
 139. Born J, Lange T, Kern W, McGregor GP, Bickel U, Fehm HL. Sniffing neuropeptides: A transnasal approach to the human brain. *Nat Neurosci* 2002;**5**:514–516.
 140. Di Simplicio M, Massey-Chase R, Cowen PJ, Harmer CJ. Oxytocin enhances processing of positive versus negative emotional information in healthy male volunteers. *J Psychopharmacol* 2009;**23**:241–248.
 141. Basson R, Berman J, Burnett A, et al. Report of the international consensus development conference on female sexual dysfunction: Definitions and classifications. *J Urol* 2000;**163**:888–893.
 142. Pfaus JG. Pathways of sexual desire. *J Sexual Med* 2009;**6**:1506–1533.
 143. McMurray G, Casey JH, Naylor AM. Animal models in urological disease and sexual dysfunction. *Br J Pharmacol* 2006;**147**:S62–S79.
 144. Flanagan LM, Pfaus JG, Pfaff DW, Mcewen BS. Induction of Fos immunoreactivity in oxytocin neurons after sexual-activity in female rats. *Neuroendocrinology* 1993;**58**:352–358.
 145. Pfaus JG, Jakob A, Kleopoulos SP, Gibbs RB, Pfaff DW. Sexual stimulation induces Fos immunoreactivity with in GnRH neurons of the female rat preoptic area—Interaction with steroid-hormones. *Neuroendocrinology* 1994;**60**:283–290.
 146. Pfaus JG, Heeb MM. Implications of immediate-early gene induction in the brain following sexual stimulation of female and male rodents. *Brain Res Bull* 1997;**44**:397–407.
 147. Blaicher W, Gruber D, Bieglmayer C, Blaicher AM, Knogler W, Huber JC. The role of oxytocin in relation to female sexual arousal. *Gynecol Obstetr Invest* 1999;**47**:125–126.
 148. Benelli A, Poggioli R, Luppi P, Ruini L, Bertolini A, Arletti R. Oxytocin enhances, and oxytocin antagonism decreases, sexual receptivity in intact female rats. *Neuropeptides* 1994;**27**:245–250.
 149. Caldwell JD, Jirikowski GF, Greer ER, Pedersen CA. Medial preoptic area oxytocin and female sexual receptivity. *Behav Neurosci* 1989;**103**:655–662.
 150. Schulze HG, Gorzalka BB. Oxytocin effects on lordosis frequency and lordosis duration following infusion into the medial pre-optic area and ventromedial hypothalamus of female rats. *Neuropeptides* 1991;**18**:99–106.
 151. Eliasson M, Meyerson BJ. Comparison of action of lysergic-acid diethylamide and apomorphine on copulatory response in female rat. *Psychopharmacology* 1976;**49**:301–306.
 152. Everitt BJ, Fuxe K, Hokfelt T, Jonsson G. Role of monoamines in control by hormones of sexual receptivity in female rat. *J Comp Physiol Psychol* 1975;**89**:556–572.
 153. Foreman MM, Hall JL. Effects of D2-dopaminergic receptor stimulation on the lordotic response of female rats. *Psychopharmacology* 1987;**91**:96–100.
 154. Hamburgerbar R, Rieger H. Apomorphine—Facilitation of sexual-behavior in female rats. *Eur J Pharmacol* 1975;**32**:357–360.
 155. Grierson JP, James MD, Pearson JR, Wilson CA. The effect of selective D1 and D2 dopaminergic agents on sexual receptivity in the female rat. *Neuropharmacology* 1988;**27**:181–189.
 156. Quysner A, Blaustein JD. A dopamine antagonist blocks vaginocervical stimulation-induced neuronal responses in the rat forebrain. *Brain Res* 2001;**921**:173–182.
 157. Meisel RL, Camp DM, Robinson TE. A microdialysis study of ventral striatal dopamine during sexual-behavior in female syrian-hamsters. *Behav Brain Res* 1993;**55**:151–157.
 158. Pfaus JG, Damsma G, Wenkstern D, Fibiger HC. Sexual-activity increases dopamine transmission in the nucleus-accumbens and striatum of female rats. *Brain Res* 1995;**693**:21–30.
 159. Foreman MM, Moss RL. Role of hypothalamic dopaminergic receptors in the control of lordosis behavior in the female rat. *Physiol Behav* 1979;**22**:283–289.
 160. Diamond LE, Earle DC, Heiman JR, Rosen RC, Perelman MA, Harning R. An effect on the subjective sexual response in premenopausal women with sexual arousal disorder by bremelanotide (PT-141), a melanocortin receptor agonist. *J Sexual Med* 2006;**3**:628–638.
 161. Shadiack AM, Sharma SD, Earle DC, Spana C, Hallam TJ. Melanocortins in the treatment of male and female sexual dysfunction. *Curr Topics Med Chem* 2007;**7**:1137–1144.
 162. Gonzaga GC, Turner RA, Keltner D, Campos B, Altemus M. Romantic love and sexual desire in close relationships. *Emotion* 2006;**6**:163–179.
 163. Nishimori K, Young LJ, Guo QX, Wang ZX, Insel TR, Matzuk MM. Oxytocin is required for nursing but is not essential for parturition or reproductive behavior. *Proc Natl Acad Sci USA* 1996;**93**:11699–11704.

164. Winslow JT, Hearn EF, Ferguson J, Young LJ, Matzuk MM, Insel TR. Infant vocalization, adult aggression, and fear behavior of an oxytocin null mutant mouse. *Horm Behav* 2000;**37**:145–155.
165. Takayanagi Y, Yoshida M, Bielsky IF, et al. Pervasive social deficits, but normal parturition, in oxytocin receptor-deficient mice. *Proc Natl Acad Sci USA* 2005;**102**:16096–16101.
166. Douglas AJ. The vasopressin and oxytocin systems. In: Steckler T, Kalin NH, Reul JM, editors. *The handbook of stress and the brain. Part 1: The neurobiology of stress. Techniques in behavioural and neural sciences series.* Amsterdam: Elsevier Science BV, 2005;205–209.
167. Rodriguiz RM, Chu R, Caron MG, Wetsel WC. Aberrant responses in social interaction of dopamine transporter knockout mice. *Behav Brain Res* 2004;**148**:185–198.
168. Tanda K, Nishi A, Matsuo N, et al. Abnormal social behaviour, hyperactivity, impaired remote spatial memory, and increased D1-mediated dopaminergic signaling in neuronal nitric oxide synthase knockout mice. *Mol Brain* 2009;**2**:19.
169. Insel TR, Young LJ. The neurobiology of attachment. *Nat Rev Neurosci* 2001;**2**:129–136.
170. Sanchez-Andrade G, Kendrick KM. The main olfactory system and social learning in mammals. *Behav Brain Res* 2009;**200**:323–335.
171. Savaskan E, Ehrhardt R, Schulz A, Walter M, Schachinger H. Post-learning intranasal oxytocin modulates human memory for facial identity. *Psychoneuroendocrinology* 2008;**33**:368–374.
172. Macbeth AH, Scharfman HE, MacLusk NJ, Gautreaux C, Luine VN. Effects of multiparity on recognition memory, monoaminergic neurotransmitters, and brain-derived neurotrophic factor (BDNF). *Horm Behav* 2008;**54**:7–17.
173. Guevara-Guzman R, Arriaga V, Kendrick KM, Bernal C, Vega X, Mercado-Gomez OF, Rivas-Arancibia S. Estradiol prevents ozone-induced increases in brain lipid peroxidation and impaired social recognition memory in female rats. *Neuroscience* 2009;**159**:940–950.
174. Insel TR, Young LJ. The neurobiology of attachment. *Nat Rev Neurosci* 2001;**2**:129–136.
175. Wersinger SR, Temple JL, Caldwell HK, Young WS. Inactivation of the oxytocin and the vasopressin (Avp) 1b receptor genes, but not the Avp 1a receptor gene, differentially impairs the Bruce effect in laboratory mice (*Mus musculus*). *Endocrinology* 2008;**149**:116–121.
176. Hammock EAD, Young LJ. Oxytocin, vasopressin and pair bonding: Implications for autism. *Philos Trans Roy Soc B-Biol Sci* 2006;**361**:2187–2198.
177. Everitt BJ, Thomas KL, Phillips GD, Morrison CH, Rowe ES. Differential induction of C-FOS in limbic and hypothalamic structures by primary and secondary reinforcers. *Eur J Neurosci* 1998;**10**:157.
178. Insel TR, Young LJ. The neurobiology of attachment. *Nat Rev Neurosci* 2001;**2**:129–136.
179. Bartels A, Zeki S. The chronoarchitecture of the human brain—Natural viewing conditions reveal a time-based anatomy of the brain. *Neuroimage* 2004;**22**:419–433.
180. Loup F, Tribollet E, Duboisdauphin M, Dreifuss JJ. Localization of high-affinity binding-sites for oxytocin and vasopressin in the human brain—An autoradiographic study. *Brain Res* 1991;**555**:220–232.
181. Uvnäs-Moberg K, Alster P, Svensson TH. Amperozide and clozapine but not haloperidol or raclopride increase the secretion of oxytocin in rats. *Psychopharmacology* 1992;**109**:473–476.
182. Heinrichs M, von Dawans B, Domes G. Oxytocin, vasopressin, and human social behavior. *Front Neuroendocrinol* 2009;**30**:548–557.
183. Kanner L. Autistic disturbances of affective contact. *Nervous Child* 1943;**2**:217–250.
184. Modahl C, Green L, Fein D, Morris M, Waterhouse L, Feinstein C, Levin H. Plasma oxytocin levels in autistic children. *Biol Psychiatry* 1998;**43**:270–277.
185. Goldman M, Marlow-O'Connor M, Torres I, Carter CS. Diminished plasma oxytocin in schizophrenic patients with neuroendocrine dysfunction and emotional deficits. *Schizophrenia Res* 2008;**98**:247–255.
186. Kirsch P, Esslinger C, Chen Q, et al. Oxytocin modulates neural circuitry for social cognition and fear in humans. *J Neurosci* 2005;**25**:11489–11493.
187. Hollander E, Bartz J, Chaplin W, et al. Oxytocin increases retention of social cognition in autism. *Biol Psychiatry* 2007;**61**:498–503.
188. Baumgartner T, Heinrichs M, Vonlanthen A, Fischbacher U, Fehr E. Oxytocin shapes the neural circuitry of trust and trust adaptation in humans. *Neuron* 2008;**58**:639–650.
189. Yrigollen CM, Han SS, Kochetkova A, et al. Genes controlling affiliative behavior as candidate genes for autism. *Biol Psychiatry* 2008;**63**:911–916.
190. Jacob S, Brune CW, Carter CS, Leventhal BL, Lord C, Cook EH. Association of the oxytocin receptor gene (OXTR) in Caucasian children and adolescents with autism. *Neurosci Lett* 2007;**417**:6–9.
191. Strathearn L. The elusive etiology of autism: Nature and nurture? *Front Behav Neurosci* 2009;**3**:1–3.
192. Meyer-Lindenberg A, Kolachana B, Gold B, et al. Genetic variants in AVPR1A linked to autism predict amygdala activation and personality traits in healthy humans. *Mol Psychiatry* 2009;**14**:968–975.
193. Gadow KD, Roohi J, DeVincent CJ, Hatchwell E. Association of ADHD, tics, and anxiety with dopamine transporter (DAT1) genotype in autism spectrum disorder. *J Child Psychol Psychiatry* 2008;**49**:1331–1338.
194. Anderson BM, Schnetz-Boutaud N, Bartlett J, et al. Examination of association to autism of common genetic variation in genes related to dopamine. *Autism Res* 2008;**1**:364–369.

195. Sun X, Yue J, Zheng C. Study of dopamine transporter imaging on the brain of children with autism. *J Biomed Eng* 2008;**25**:327–330.
196. Sharp SL, McQuillin A, Gurling HMD. Genetics of attention-deficit hyperactivity disorder (ADHD). *Neuropharmacology* 2009;**57**:590–600.
197. Schnetz-Boutaud NC, Anderson BM, Brown KD, et al. Examination of tetrahydrobiopterin pathway genes in autism. *Genes Brain Behav* 2009;**8**:753–757.
198. Hollander E, Novotny S, Hanratty M, Yaffe R, DeCaria CM, Aronowitz BR, Mosovich S. Oxytocin infusion reduces repetitive behaviors in adults with autistic and Asperger's disorders. *Neuropsychopharmacology* 2003;**28**:193–198.
199. Kosfeld M, Heinrichs M, Zak PJ, Fischbacher U, Fehr E. Oxytocin increases trust in humans. *Nature* 2005;**435**:673–676.
200. Herguner S, Mukaddes NM. Risperidone-induced enuresis in two children with autistic disorder. *J Child Adolesc Psychopharmacol* 2007;**17**:527–530.
201. Leng G, Meddle SL, Douglas AJ. Oxytocin and the maternal brain. *Curr Opin Pharmacol* 2008;**8**:731–734.
202. Young LJ, Winslow JT, Wang ZX, Gingrich B, Guo QX, Matzuk MM, Insel TR. Gene targeting approaches to neuroendocrinology: Oxytocin, maternal behavior, and affiliation. *Horm Behav* 1997;**31**:221–231.
203. Lonstein JS. Effects of dopamine receptor antagonism with haloperidol on nurturing behavior in the biparental prairie vole. *Pharmacol Biochem Behav* 2002;**74**:11–19.
204. Bales KL, Kim AJ, Lewis-Reese AD, Carter CS. Both oxytocin and vasopressin may influence alloparental behavior in male prairie voles. *Horm Behav* 2004;**45**:354–361.
205. Wynne-Edwards KE, Timonin ME. Paternal care in rodents: Weakening support for hormonal regulation of the transition to behavioral fatherhood in rodent animal models of biparental care. *Horm Behav* 2007;**52**:114–121.
206. de Jong TR, Chauke M, Harris BN, Saltzman W. From here to paternity: Neural correlates of the onset of paternal behavior in California mice (*Peromyscus californicus*). *Horm Behav* 2009;**56**:220–231.
207. Kinsley CH, Lambert KG. Reproduction-induced neuroplasticity: Natural behavioural and neuronal alterations associated with the production and care of offspring. *J Neuroendocrinol* 2008;**20**:515–525.
208. Lonstein JS. Regulation of anxiety during the postpartum period. *Front Neuroendocrinol* 2007;**28**:115–141.
209. Ross HE, Young LJ. Oxytocin and the neural mechanisms regulating social cognition and affiliative behaviour. *Front Neuroendocrinol* 2009;**30**:534–547.
210. Nissen E, Gustavsson P, Widstrom AM, Uvnas-Moberg K. Oxytocin, prolactin, milk production and their relationship with personality traits in women after vaginal delivery or Cesarean section. *J Psychosomat Obstetr Gynecol* 1998;**19**:49–58.
211. Francis DD, Young LJ, Meaney MJ, Insel TR. Naturally occurring differences in maternal care are associated with the expression of oxytocin and vasopressin (V1a) receptors: Gender differences. *J Neuroendocrinol* 2002;**14**:349–353.
212. Champagne FA, Meaney MJ. Trans generational effects of social environment on variations in maternal care and behavioral response to novelty. *Behav Neurosci* 2007;**121**:1353–1363.
213. Champagne FA, Meaney MJ. Stress during gestation alters postpartum maternal care and the development of the offspring in a rodent model. *Biol Psychiatry* 2006;**59**:1227–1235.
214. Bartels A, Zeki S. The neural correlates of maternal and romantic love. *Neuroimage* 2004;**21**:1155–1166.
215. Uvnas-Moberg K. Oxytocin may mediate the benefits of positive social interaction and emotions. *Psychoneuroendocrinology* 1998;**23**:819–835.
216. Holman SD, Goy RW. Experiential and hormonal correlates of caregiving in rhesus monkeys. In: Pryce CR, Martin RD, editors. *Motherhood in human and non-human primates, biosocial determinants*. Basel: Karger, 1995;87–93.
217. Spieglewsky C, Roubert C, Hamon M, Nosten-Bertrand M, Betancur C, Giros B. Behavioural disturbances associated with hyperdopaminergia in dopamine-transporter knockout mice. *Behav Pharmacol* 2000;**11**:279–290.
218. Kavelaars A, Cobelens PM, Teunis MAT, Heijnen CJ. Changes in innate and acquired immune responses in mice with targeted deletion of the dopamine transporter gene. *J Neuroimmunol* 2005;**161**:162–168.
219. Champagne FA, Chretien P, Stevenson CW, Zhang TY, Gratton A, Meaney MJ. Variations in nucleus accumbens dopamine associated with individual differences in maternal behavior in the rat. *J Neurosci* 2004;**24**:4113–4123.
220. Shakrok H, Meaney MJ. Oxytocin-dopamine interactions mediate variations in maternal behavior in the rat. *Endocrinology* 2010;**151**:1–11.
221. Sullivan RM. Developing a sense of safety—The neurobiology of neonatal attachment. *Ann N Y Acad Sci* 2003;**1008**:122–131.
222. Seckl JR, Meaney MJ. Glucocorticoid programming. *Ann N Y Acad Sci* 2004;**1032**:63–84.
223. McArthur S, McHale E, Gillies GE. The size and distribution of midbrain dopaminergic populations are permanently altered by perinatal glucocorticoid exposure in a sex- region- and time-specific manner. *Neuropsychopharmacology* 2007;**32**:1462–1476.
224. Engert V, Joobar R, Meaney MJ, Hellhammer DH, Pruessner JC. Behavioral response to methylphenidate challenge: Influence of early life parental care. *Develop Psychobiol* 2009;**51**:408–416.

225. Bakermans-Kranenburg MJ, van IJzendoorn MH. Oxytocin receptor (OXTR) and serotonin transporter (5-HTT) genes associated with observed parenting. *Cogn Affect Neurosci* 2008;**3**:128–134.
226. Clemens KJ, Cornish JL, Hunt GE, McGregor LS. Repeated weekly exposure to MDMA, methamphetamine or their combination: Long-term behavioural and neurochemical effects in rats. *Drug Alcohol Depend* 2007;**86**:183–190.
227. Miczek KA, Covington HE, Nikulina EA, Hammer RP. Aggression and defeat: Persistent effects on cocaine self-administration and gene expression in peptidergic and aminergic mesocorticolimbic circuits. *Neurosci Biobehav Rev* 2004;**27**:787–802.
228. Brown CH, Russell JA. Cellular mechanisms underlying neuronal excitability during morphine withdrawal in physical dependence: Lessons from the magnocellular oxytocin system. *Stress-Int J Biol Stress* 2004;**7**: 97–107.
229. Sarnyai Z, Kovacs GL. Role of oxytocin in the neuroadaptation to drugs of abuse. *Psychoneuroendocrinology* 1994;**19**:85–117.
230. Galloway MP. Neurochemical interactions of cocaine with dopaminergic systems. *Trends Pharmacol Sci* 1988;**9**:451–454.
231. Koob GF, Hubner CB. Reinforcement pathways for cocaine. *NIDA Res Monogr* 1988;**88**:137–159.
232. Koob GF, Swerdlow NR. The functional output of the mesolimbic dopamine system. *Ann N Y Acad Sci* 1988;**537**:216–227.
233. Koob GF, Sanna PP, Bloom FE. Neuroscience of addiction. *Neuron* 1998;**21**:467–476.
234. McGregor IS, Callaghan PD, Hunt GE. From ultrasocial to antisocial: A role for oxytocin in the acute reinforcing effects and long-term adverse consequences of drug use? *Br J Pharmacol* 2008;**154**:358–368.
235. Morley KC, Gallate JE, Hunt GE, Mallet PE, McGregor IS. Increased anxiety and impaired memory in rats 3 months after administration of 3,4-methylenedioxymethamphetamine (“Ecstasy”). *Eur J Pharmacol* 2001;**433**:91–99.
236. Blatchford KE, Diamond K, Westbrook RF, McNally GP. Increased vulnerability to stress following opiate exposures: Behavioral and autonomic correlates. *Behav Neurosci* 2005;**119**:1034–1041.
237. Breiter HC, Gollub RL, Weisskoff RM, et al. Acute effects of cocaine on human brain activity and emotion. *Neuron* 1997;**19**:591–611.
238. Wozniak KM, Pert A, Mele A, Linnoila M. Focal application of alcohols elevates extracellular dopamine in rat-brain—A microdialysis study. *Brain Res* 1991;**540**:31–40.
239. Pettit HO, Justice JB. Effect of dose on cocaine self-administration behavior and dopamine levels in the nucleus-accumbens. *Brain Res* 1991;**539**:94–102.
240. Schmidt HD, Pierce RC. Cooperative activation of D1-like and D2-like dopamine receptors in the nucleus accumbens shell is required for the reinstatement of cocaine-seeking behavior in the rat. *Neuroscience* 2006;**142**:451–461.
241. Bari AA, Pierce RC. D1-like and D2 dopamine receptor antagonists administered into the shell subregion of the rat nucleus accumbens decrease cocaine, but not food, reinforcement. *Neuroscience* 2005;**135**:959–968.
242. Caine SB, Koob GF. Effects of mesolimbic dopamine depletion on responding maintained by cocaine and food. *J Exp Anal Behav* 1994;**61**:213–221.
243. Vanleeuwen FW, Vanheerikhuizen J, Vandermeulen G, Wolters P. Light microscopic autoradiographic localization of [H-3] oxytocin binding-sites in the rat-brain, pituitary and mammary-gland. *Brain Res* 1985;**359**:320–325.
244. Sarnyai Z, Vecsernyes M, Laczi F, Biro E, Szabo G, Kovacs GL. Effects of cocaine on the contents of neurohypophyseal hormones in the plasma and in different brain structures in rats. *Neuropeptides* 1992;**23**:27–31.
245. Kovacs GL, Sarnyai Z, Babarczy E, Szabo G, Telegdy G. The role of oxytocin dopamine interactions in cocaine-induced locomotor hyperactivity. *Neuropharmacology* 1990;**29**:365–368.
246. Qi J, Yang JY, Song M, Li Y, Wang F, Wu CF. Inhibition by oxytocin of methamphetamine-induced hyperactivity related to dopamine turnover in the mesolimbic region in mice. *Naunyn-Schmiedeberg Arch Pharmacol* 2008;**376**:441–448.
247. Kovacs GL, Sarnyai Z, Babarczy E, Szabo G, Telegdy G. The role of oxytocin dopamine interactions in cocaine-induced locomotor hyperactivity. *Neuropharmacology* 1990;**29**:365–368.
248. Sarnyai Z. Oxytocin and neuroadaptation to cocaine. *Adv Brain Vasopressin* 1998;**119**:449–466.
249. Sarnyai Z, Kovacs GL. Role of oxytocin in the neuroadaptation to drugs of abuse. *Psychoneuroendocrinology* 1994;**19**:85–117.
250. Cui SS, Bowen RC, Gu GB, Hannesson DK, Yu PH, Zhang X. Prevention of cannabinoid withdrawal syndrome by lithium: Involvement of oxytocinergic neuronal activation. *J Neurosci* 2001;**21**:9867–9876.
251. Russell JA, Leng G, Bicknell RJ. Opioid tolerance and dependence in the magnocellular oxytocin system—A physiological mechanism. *Exp Physiol* 1995;**80**:307–340.
252. Diana M, Pistis M, Carboni S, Gessa GL, Rossetti ZL. Profound decrement of mesolimbic dopaminergic neuronal-activity during ethanol withdrawal syndrome in rats—Electrophysiological and biochemical-evidence. *Proc Natl Acad Sci USA* 1993;**90**:7966–7969.
253. Weiss F, Parsons LH, Schulteis G, Hyytia P, Lorang MT, Bloom FE, Koob GF. Ethanol self-administration restores withdrawal-associated deficiencies in accumbal

- dopamine and 5-hydroxytryptamine release in dependent rats. *J Neurosci* 1996;**16**:3474–3485.
254. Slemmer JE, Martin BR, Damaj MI. Bupropion is a nicotinic antagonist. *J Pharmacol Exp Therap* 2000;**295**:321–327.
 255. Cornuz J. Smoking cessation interventions in clinical practice. *Eur J Vasc Endovasc Surg* 2007;**34**:397–404.
 256. Eisenberg MJ, Filion KB, Yavin D, et al. Pharmacotherapies for smoking cessation: A meta-analysis of randomized controlled trials. *Can Med Assoc J* 2008;**179**:135–144.
 257. Yahyavi-Firouz-Abadi N, See RE. Anti-relapse medications: Preclinical models for drug addiction treatment. *Pharmacol Therap* 2009;**124**:235–247.
 258. Haney M, Hart CL, Foltin RW. Effects of bupropion on cocaine self-administration: Opioid- and nonopioid-dependent volunteers. *Neuropsychopharmacology* 2006;**31**:1814–1821.
 259. Johnson BA. Recent advances in the development of treatments for alcohol and cocaine dependence: Focus on topiramate and other modulators of GABA or glutamate function. *CNS Drugs* 2005;**19**:873–896.
 260. Shoptaw S, Yang XW, Rotheram-Fuller EJ, Hsieh YCM, Kintaudi PC, Charuvastra VC, Ling W. Randomized placebo-controlled trial of bupropion for cocaine dependence: Preliminary effects for individuals with chronic patterns of cocaine use. *J Clin Psychiatry* 2003;**64**:1440–1448.
 261. Poling J, Oliveto A, Petry N, et al. Six-month trial of Bupropion with contingency management for cocaine dependence in a methadone-maintained population. *Arch Gen Psychiatry* 2006;**63**:219–228.
 262. Mattick RP, Breen C, Kimber J, Davoli M. Methadone maintenance therapy versus no opioid replacement therapy for opioid dependence. *Cochrane Database Syst Rev* 2009;**8**:1–31.
 263. Carroll KM, Nich C, Ball SA, McCance E, Rounsaville BJ. Treatment of cocaine and alcohol dependence with psychotherapy and disulfiram. *Addiction* 1998;**93**:713–727.
 264. Gonzalez G, Sevarino K, Sofuoglu M, et al. Tiagabine increases cocaine-free urines in cocaine-dependent methadone-treated patients: Results of a randomized pilot study. *Addiction* 2003;**98**:1625–1632.
 265. Silva SM, Madeira MD, Ruela C, Paula-Barbosa MM. Prolonged alcohol intake leads to irreversible loss of vasopressin and oxytocin neurons in the paraventricular nucleus of the hypothalamus. *Brain Res* 2002;**925**:76–88.
 266. Kaye W. Neurobiology of anorexia and bulimia nervosa. *Physiol Behav* 2008;**94**:121–135.
 267. Johnstone LE, Fong TM, Leng G. Neuronal activation in the hypothalamus and brainstem during feeding in rats. *Cell Metabol* 2006;**4**:313–321.
 268. Douglas AJ, Johnstone LE, Leng G. Neuroendocrine mechanisms of change in food intake during pregnancy: A potential role for brain oxytocin. *Physiol Behav* 2007;**91**:352–365.
 269. Douglas AJ, Johnstone LE, Leng G. Neuroendocrine mechanisms of change in food intake during pregnancy: A potential role for brain oxytocin. *Physiol Behav* 2007;**91**:352–365.
 270. Leng G, Caquineau C. Oxytocin control of food intake. In: Preedy VR, editor. *The handbook of behaviour, diet and nutrition*. New York: Springer, 2009.
 271. Fetissov SO, Harro J, Jaanisk M, et al. Autoantibodies against neuropeptides are associated with psychological traits in eating disorders. *Proc Natl Acad Sci USA* 2005;**102**:14865–14870.
 272. Shimizu H, Ohsaki A, Oh I, Okada S, Mori M. A new anorexigenic protein, nesfatin-1. *Peptides* 2009;**30**:995–998.
 273. Maejima Y, Sedbazar U, Suyama S, et al. Nesfatin-1-regulated oxytocinergic signaling in the paraventricular nucleus causes anorexia through a leptin-independent melanocortin pathway. *Cell Metab* 2009;**10**:355–365.
 274. Swaab DF. Neuropeptides in hypothalamic neuronal disorders. *Int Rev Cytol—A Surv Cell Biol* 2004;**240**:305–375.
 275. Wirth MM, Olszewski PK, Levine AS, Giraudo SQ. Effect of Agouti-related protein on development of conditioned taste aversion and oxytocin neuronal activation. *Neuroreport* 2002;**13**:1355–1358.
 276. Wise RA. Role of brain dopamine in food reward and reinforcement. *Philos Trans R Soc Lond B Biol Sci* 2006;**361**:1149–1158.
 277. Abizaid A, Horvath TL. Brain circuits regulating energy homeostasis. *Regulat Peptides* 2008;**149**:3–10.
 278. Narayanan NS, Guarnieri DJ, DiLeone RJ. Metabolic hormones, dopamine circuits, and feeding. *Front Neuroendocrinol* 2010;**31**:104–112.
 279. Naleid AM, Grace MK, Cummings DE, Levine AS. Ghrelin induces feeding in the mesolimbic reward pathway between the ventral tegmental area and the nucleus accumbens. *Peptides* 2005;**26**:2274–2279.
 280. Fraley GS, Dinh TT, Ritter S. Immunotoxic catecholamine lesions attenuate 2DG-induced increase of AGRP mRNA. *Peptides* 2002;**23**:1093–1099.
 281. Kruger S, Kennedy SH. Psychopharmacotherapy of anorexia nervosa, bulimia nervosa and binge-eating disorder. *J Psychiatry Neurosci* 2000;**25**:497–508.
 282. Shinohara M, Mizushima H, Hirano M, et al. Eating disorders with binge-eating behaviour are associated with the s allele of the 3'-UTR VNTR polymorphism of the dopamine transporter gene. *J Psychiatry Neurosci* 2004;**29**:134–137.
 283. Modell S, Holsboer F. Depression and effects of antidepressant drugs on the stress systems. In: Steckler T, Kalin NH, Reul JMHM, editors, *Handbook of stress and the brain*. Amsterdam: Elsevier, 2005;273–286.

284. Vonbardeleben U, Holsboer F. Effect of age on the cortisol response to human corticotropin-releasing hormone in depressed-patients pretreated with dexamethasone. *Biol Psychiatry* 1991;**29**:1042–1050.
285. Mizoguchi K, Shoji H, Ikeda R, Tanaka Y, Tabira T. Persistent depressive state after chronic stress in rats is accompanied by HPA axis dysregulation and reduced prefrontal dopaminergic neurotransmission. *Pharmacol Biochem Behav* 2008;**91**:170–175.
286. Ingram CD. Pathways and transmitter interactions mediating an integrated stress response. In: Steckler T, Kalin NH, Reul JM, editors. *Techniques in the behavioural and neural sciences series*. Amsterdam: Elsevier Science BV, 2005;609–642.
287. Kim S, Webster MJ. The Stanley neuropathology consortium integrative database: A novel, web-based tool for exploring neuropathological markers in psychiatric disorders and the biological processes associated with abnormalities of those markers. *Neuropsychopharmacology* 2010;**35**:473–482.
288. Mizoguchi K, Shoji H, Ikeda R, Tanaka Y, Tabira T. Persistent depressive state after chronic stress in rats is accompanied by HPA axis dysregulation and reduced prefrontal dopaminergic neurotransmission. *Pharmacol Biochem Behav* 2008;**91**:170–175.
289. Seo DJ, Patrick CJ, Kennealy PJ. Role of serotonin and dopamine system interactions in the neurobiology of impulsive aggression and its comorbidity with other clinical disorders. *Aggress Violent Behav* 2008;**13**:383–395.
290. Nikolaus S, Antke C, Muller HW. In vivo imaging of synaptic function in the central nervous system: II. Mental and affective disorders. *Behav Brain Res* 2009;**204**:32–66.
291. Yoshida M, Takayanagi Y, Inoue K, Kimura T, Young LJ, Onaka T, Nishimori K. Evidence that oxytocin exerts anxiolytic effects via oxytocin receptor expressed in serotonergic neurons in mice. *J Neurosci* 2009;**29**:2259–2271.
292. Di S, Malcher-Lopes R, Marcheselli VL, Bazan NG, Tasker JG. Rapid glucocorticoid-mediated endocannabinoid release and opposing regulation of glutamate and gamma-aminobutyric acid inputs to hypothalamic magnocellular neurons. *Endocrinology* 2005;**146**:4292–4301.
293. Ozsoy S, Esel E, Kula M. Serum oxytocin levels in patients with depression and the effects of gender and antidepressant treatment. *Psychiatry Res* 2009;**169**:249–252.
294. Neumann ID. Brain oxytocin: A key regulator of emotional and social behaviours in both females and males. *J Neuroendocrinol* 2008;**20**:858–865.
295. Brunton PJ, Russell JA. Keeping oxytocin neurons under control during stress in pregnancy. *Prog Brain Res* 2008;**170**:365–377.
296. Boccia ML, Pedersen CA. Brief vs. long maternal separations in infancy: Contrasting relationships with adult maternal behavior and lactation levels of aggression and anxiety. *Psychoneuroendocrinology* 2001;**26**:657–672.
297. Scantamburlo G, Hansenne M, Fuchs S, et al. AVP- and OT-neurophysins response to apomorphine and clonidine in major depression. *Psychoneuroendocrinology* 2005;**30**:839–845.
298. Guiard BP, El Mansari M, Blier P. Prospect of a dopamine contribution in the next generation of antidepressant drugs: The triple reuptake inhibitors. *Curr Drug Target* 2009;**10**:1069–1084.
299. Koenig AM, Thase ME. First-line pharmacotherapies for depression—What is the best choice? *Polskie Archiwum Medycyny Wewnętrznej-Polish ArchInter Med* 2009;**119**:478–485.
300. Breuer ME, Groenink L, Oosting RS, Buerger E, Korte M, Ferger B, Olivier B. Antidepressant effects of pramipexole, a dopamine D-3/D-2 receptor agonist, and 7-OH-DPAT, a dopamine D-3 receptor agonist, in olfactory bulbectomized rats. *Eur J Pharmacol* 2009;**616**:134–140.
301. Hughes S, Cohen D. A systematic review of long-term studies of drug treated and non-drug treated depression. *J Affect Disord* 2009;**118**:9–18.